Three-dimensional post-glacial expansion and diversification of an exploited oceanic fish
Shum, P, Pampoulie, C, Kristinsson, K and Mariani, S
http://dx.doi.org/10.1111/mec.13262

<table>
<thead>
<tr>
<th>Title</th>
<th>Three-dimensional post-glacial expansion and diversification of an exploited oceanic fish</th>
</tr>
</thead>
<tbody>
<tr>
<td>Authors</td>
<td>Shum, P, Pampoulie, C, Kristinsson, K and Mariani, S</td>
</tr>
<tr>
<td>Type</td>
<td>Article</td>
</tr>
<tr>
<td>URL</td>
<td>This version is available at: <a href="http://usir.salford.ac.uk/35233/">http://usir.salford.ac.uk/35233/</a></td>
</tr>
<tr>
<td>Published Date</td>
<td>2015</td>
</tr>
</tbody>
</table>

USIR is a digital collection of the research output of the University of Salford. Where copyright permits, full text material held in the repository is made freely available online and can be read, downloaded and copied for non-commercial private study or research purposes. Please check the manuscript for any further copyright restrictions.

For more information, including our policy and submission procedure, please contact the Repository Team at: usir@salford.ac.uk.
Three-dimensional post-glacial expansion and diversification of an exploited oceanic fish

PETER SHUM,* CHRISTOPHE PAMPOULIE,† KRISTJÁN KRISTINSSON† and STEFANO MARIANI*

*Ecosystems and Environment Research Centre, School of Environment and Life Sciences, University of Salford, Salford M5 4WT, UK, †Marine Research Institute, Skúlagata 4, 101 Reykjavík, Iceland

Abstract
Vertical divergence in marine organisms is being increasingly documented, yet much remains to be carried out to understand the role of depth in the context of phylogeographic reconstruction and the identification of management units. An ideal study system to address this issue is the beaked redfish, Sebastes mentella – one of four species of ‘redfish’ occurring in the North Atlantic – which is known for a widely distributed ‘shallow-pelagic’ oceanic type inhabiting waters between 250 and 550 m, and a more localized ‘deep-pelagic’ population dwelling between 550 and 800 m, in the oceanic habitat of the Irminger Sea. Here, we investigate the extent of population structure in relation to both depth and geographic spread of oceanic beaked redfish throughout most of its distribution range. By sequencing the mitochondrial control region of 261 redfish collected over a decadal interval, and combining 160 rhodopsin coding nuclear sequences and previously genotyped microsatellite data, we map the existence of two strongly divergent evolutionary lineages with significantly different distribution patterns and historical demography, and whose genetic variance is mostly explained by depth. Combined genetic data, analysed via independent approaches, are consistent with a Late Pleistocene lineage split, where segregation by depth probably resulted from the interplay of climatic and oceanographic processes with life history and behavioural traits. The ongoing process of diversification in North Atlantic S. mentella may serve as an ‘hourglass’ to understand speciation and adaptive radiation in Sebastes and in other marine taxa distributed across a depth gradient.

Keywords: approximate Bayesian computation, depth, population divergence, redfish, Sebastes, speciation

Received 17 March 2015; revision received 22 May 2015; accepted 29 May 2015

Introduction
While recent studies continue to amass evidence for notable patterns of population structure in the oceans (Sala-Bozano et al. 2009; Therkildsen et al. 2013; Silva et al. 2014), the long-standing paradigm of limited geographic barriers, high dispersal potential and large effective population size continues to hold when comparing oceanic with continental environments (Hauser & Carvalho 2008). Despite interpretive frameworks that help explain marine population structure based on geographical distance (Palumbi 1994; Weersing & Toonen 2009), oceanographic features (Gaither et al. 2010; Galindo et al. 2010; Selkoe & Toonen 2011) and life histories (Hare et al. 2005; Riginos et al. 2014), the reconstruction of connectivity patterns and recent evolutionary processes in oceanic species remains a mighty task, due to the aforementioned ‘marine paradigm’ (see Waples 1998; Avise 2004; Hauser & Carvalho 2008) and the inherent difficulty – compared to terrestrial and freshwater habitats – of even locating populations and collecting representative samples over vast areas.

All of the above is further compounded by a conspicuous, yet seldom investigated, factor: depth. Depth physically multiplies the expanses across which individuals
can disperse and populations can spread, over both ecological and evolutionary timescales. Depth is also associated with a wide variety of environmental factors, including temperature, salinity, hydrostatic pressure, food resources and light, which, independently or in combination, influence physiology, behaviour and adaptations (Somero 1992; Hyde et al. 2008; Vonlanthen et al. 2008; Irwin 2012; Roy et al. 2012; Yancey et al. 2014). As more studies document the emergence of depth-related patterns, it becomes apparent that this factor plays a role in the formation of distinct biological units, even in large, near-continuously distributed populations (Dieckmann & Doebeli 2003; Vonlanthen et al. 2008; Knutsen et al. 2009; Stefánsson et al. 2009a; Ingram 2010; Roy et al. 2012; Jennings et al. 2013). Sea level fluctuations during the last glacial cycle have been shown to have a role in shaping aquatic environments (Blanchon & Shaw 1995) and are thought to have contributed to the isolation and divergence of marine organisms (Ludt & Rocha 2015). Furthermore, the distribution of oceanic species over vast geographical areas is impacted by complex, large-scale ocean circulation processes, interplaying with bathymetric features along continental margins and mid-ocean ridges (Hemmer-Hansen et al. 2007; Knutsen et al. 2009). The full understanding of the role of depth in the context of physical oceanography, complex life histories and the historical backdrop appears as one main topical challenge in marine population biology, which may also have significant consequences for conservation and management.

One taxonomical group that has played a pivotal role in shaping our understanding of adaptive divergence along depth gradients in marine environments is the genus Sebastes (Alessandrini & Bernardi 1999; Hyde & Vetter 2007; Sivasundar & Palumbi 2010). While most species of Sebastes are limited to the North Pacific (n > 100), with two occurring in the Southern Hemisphere, the four ‘young’ North Atlantic (NA) species are thought to have descended from an ancestor of the North Pacific Sebastes alutus lineage that invaded the NA under 3 my ago, when the Bering Strait opened as a result of warming Arctic waters (Raymo et al. 1990; Love et al. 2002; Hyde & Vetter 2007). Sebastes spp. are ovoviviparous (i.e. internal fertilization), long-lived, slow-growing and late-maturing species with low natural mortality; such unique life history characteristics influence complex population structuring (Cadrin et al. 2010). The beaked redfish, S. mentella, widely distributed throughout the boreal waters of the North Atlantic and Arctic Oceans, exhibits complex depth-associated patterns of substructure (Stefánsson et al. 2009a; Cadrin et al. 2010; Shum et al. 2014). While a demersal unit is recognized along the Icelandic continental shelf and slopes (Cadrin et al. 2010), S. mentella exhibits a primarily pelagic, open-ocean behaviour, occurring in assemblages at different depth layers, with a widely distributed shallow-pelagic group inhabiting depths between 200 and 550 m, and a deep-pelagic group between 550 and 800 m, mostly circumscribed to the northeast Irminger Sea (Stefánsson et al. 2009a, b; Cadrin et al. 2010; Planque et al. 2013; Shum et al. 2014).

Almost three decades of research investigating the geographic patterns and genetic structuring of S. mentella have resulted in intense debates (Cadrin et al. 2011; Makhrov et al. 2011) regarding the number of genetically distinct populations, how they are spatially structured and connected, and how and when they have achieved their current distribution. All efforts so far had been based on traditional sampling campaigns mostly focused on geographical/fishery coverage (Cadrin et al. 2010; Shum et al. 2014), with a lack of adequate standardized sampling and replication across depth layers where different beaked redfish assemblages occur (Bergstad 2013; Planque et al. 2013). Furthermore, previous studies lacked either the appropriate molecular and analytical toolkit (Stefánsson et al. 2009b) or the sampling coverage (Shum et al. 2014) to robustly test specific phylogeographic hypotheses.

Here, we present the first extensive sampling and phylogeographic investigation of pelagic S. mentella in the North Atlantic using mitochondrial and nuclear genetic markers to resolve the interplay between longitudinal, latitudinal and vertical gradients in shaping the structure of this oceanic species. Specifically, we aim to (i) examine the phylogeographic history of oceanic S. mentella across the northeast Atlantic, with special attention to the main two clades identified in Shum et al. (2014), (ii) assess the degree to which mitochondrial lineage distribution fits with the currently perceived S. mentella stock structure and (iii) reconstruct the most realistic scenario for the evolution of this species in the North Atlantic.

Materials and methods

Sample collection

Samples were collected as part of the Icelandic Marine Research Institute 2013 survey, conducted from 11th June to the 6th July 2013 throughout the Irminger Sea. Trawling for redfish involved collecting replicated samples along a survey transect from three geographical zones at seven sites including shallow (above 500 m) and deep (below 500 m) habitats (Fig. 1, Table 1). A MultiSampler unit with three separate codends was attached to the end of the pelagic trawl enabling the collection of several independent discrete samples at three different depth strata in one trawl.
haul (Engäs et al. 1997). The opening and closing of the nets were programmed to specific depths using a timing system with approximate sampling depths ranging between 200 and 350 m, 350 and 550 m, and 550 and 900 m. These replicated sampling sites were targeted to explore the genetic variance associated

Table 1 Location, group code, year of capture, trawl depth range, position, number of individuals (N), and nucleotide (π) and haplotype (h) diversities of *Sebastes mentella* investigated for 2013 and 2006/2007 data from the North Atlantic

<table>
<thead>
<tr>
<th>Location</th>
<th>Group code</th>
<th>Year</th>
<th>Gear</th>
<th>Trawl depth range (N)</th>
<th>Latitude</th>
<th>Longitude</th>
<th>N</th>
<th>π + SD</th>
<th>h + SD</th>
</tr>
</thead>
<tbody>
<tr>
<td>Irminger Sea</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Northeast deep</td>
<td>IrNE DP 13</td>
<td>2013</td>
<td>MS</td>
<td>550–650</td>
<td>62.44</td>
<td>–29.08</td>
<td>26</td>
<td>0.003 ± 0.0006</td>
<td>0.754 ± 0.090</td>
</tr>
<tr>
<td>Northeast shallow</td>
<td>IrNE DP 13</td>
<td>2013</td>
<td>MS</td>
<td>250–300</td>
<td>62.45</td>
<td>–30.21</td>
<td>11</td>
<td>0.006 ± 0.0019</td>
<td>0.927 ± 0.066</td>
</tr>
<tr>
<td>West shallow</td>
<td>IrW SP 13</td>
<td>2013</td>
<td>MS</td>
<td>200–275</td>
<td>61.00</td>
<td>–38.53</td>
<td>23</td>
<td>0.004 ± 0.0007</td>
<td>0.893 ± 0.036</td>
</tr>
<tr>
<td>West intermediate</td>
<td>IrW SP 13</td>
<td>2013</td>
<td>MS</td>
<td>475</td>
<td>61.45</td>
<td>–40.19</td>
<td>16</td>
<td>0.004 ± 0.0004</td>
<td>0.908 ± 0.040</td>
</tr>
<tr>
<td>West deep</td>
<td>IrW SP 13</td>
<td>2013</td>
<td>MS</td>
<td>600</td>
<td>61.44</td>
<td>–29.11</td>
<td>26</td>
<td>0.005 ± 0.0006</td>
<td>0.898 ± 0.029</td>
</tr>
<tr>
<td>Southwest shallow</td>
<td>IrSW SP 13</td>
<td>2013</td>
<td>MS</td>
<td>300</td>
<td>57.44</td>
<td>–45.40</td>
<td>33</td>
<td>0.004 ± 0.0005</td>
<td>0.883 ± 0.035</td>
</tr>
<tr>
<td>Southwest deep</td>
<td>IrSW DP 13</td>
<td>2013</td>
<td>MS</td>
<td>550–850</td>
<td>57.45</td>
<td>–41.04</td>
<td>8</td>
<td>0.004 ± 0.0007</td>
<td>0.857 ± 0.108</td>
</tr>
<tr>
<td>Archived samples</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Irminger Sea</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Northeast deep</td>
<td>IrNE DP 06</td>
<td>2006</td>
<td>Pe</td>
<td>284</td>
<td>58.38</td>
<td>–43</td>
<td>25</td>
<td>0.004 ± 0.0005</td>
<td>0.883 ± 0.035</td>
</tr>
<tr>
<td>Northeast deep</td>
<td>IrNE DP 06</td>
<td>2006</td>
<td>Pe</td>
<td>786</td>
<td>61.44</td>
<td>–29.11</td>
<td>26</td>
<td>0.002 ± 0.0005</td>
<td>0.578 ± 0.114</td>
</tr>
<tr>
<td>Faroe Islands west</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>East</td>
<td>FW1 DP 07</td>
<td>2007</td>
<td>Dm</td>
<td>589–644</td>
<td>60.59</td>
<td>–9.32</td>
<td>9</td>
<td>0.006 ± 0.0014</td>
<td>0.917 ± 0.092</td>
</tr>
<tr>
<td>West</td>
<td>FW2 DP 07</td>
<td>2007</td>
<td>Dm</td>
<td>704–658</td>
<td>60.44</td>
<td>–9.57</td>
<td>11</td>
<td>0.006 ± 0.0008</td>
<td>0.927 ± 0.066</td>
</tr>
<tr>
<td>Faroe Islands east</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>South</td>
<td>FE1 DP 07</td>
<td>2007</td>
<td>Dm</td>
<td>388–401</td>
<td>61.59</td>
<td>–4.19</td>
<td>9</td>
<td>0.007 ± 0.0027</td>
<td>0.833 ± 0.127</td>
</tr>
<tr>
<td>North</td>
<td>FE2 DP 07</td>
<td>2007</td>
<td>Dm</td>
<td>366–433</td>
<td>62.21</td>
<td>–4.13</td>
<td>9</td>
<td>0.005 ± 0.0009</td>
<td>0.911 ± 0.077</td>
</tr>
<tr>
<td>Norwegian shelf</td>
<td>NS SP 07</td>
<td>2007</td>
<td>Dm</td>
<td>430</td>
<td>66.14</td>
<td>6.2</td>
<td>10</td>
<td>0.002 ± 0.0008</td>
<td>0.533 ± 0.180</td>
</tr>
<tr>
<td>Norwegian international waters</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>South</td>
<td>NIW1 SP 07</td>
<td>2006</td>
<td>Pe</td>
<td>440</td>
<td>72.01</td>
<td>5.3</td>
<td>10</td>
<td>0.003 ± 0.0010</td>
<td>0.800 ± 0.100</td>
</tr>
<tr>
<td>North</td>
<td>NIW2 SP 07</td>
<td>2006</td>
<td>Pe</td>
<td>445</td>
<td>72.21</td>
<td>5.15</td>
<td>10</td>
<td>0.003 ± 0.0009</td>
<td>0.822 ± 0.097</td>
</tr>
</tbody>
</table>

MS, multisampler; Dm, demersal; Pe, pelagic trawls.

© 2015 The Authors. *Molecular Ecology* Published by & Sons Ltd.
with geography (latitude, longitude) and depth for *Sebastes mentella*.

Between five and 30 specimens were collected and sampled from each haul section, for a total of 143 dorsal fin clip tissue samples of *S. mentella* collected at seven sites distributed throughout the Irminger Sea and stored in 100% ethanol. The data set was also complemented by the 50 mtDNA control region and 22 rhodopsin sequences that first revealed the existence of two depth-associated *S. mentella* clades in the Irminger Sea (Shum et al. 2014; GenBank Accession nos: mtDNA, KM013849-KM013898; rhodopsin, KM013899-KM013920.1) and by 68 randomly selected archived tissue samples (previously genotyped at 12 microsatellite loci by Stefansson et al. 2009a), to better cover the current biogeographical spread of the species: west Faroe (n = 20) and east Faroe (n = 18) collected off the Faroese shelf in 2007, Norwegian shelf (n = 10), and pelagic collections from Norwegian international waters (n = 20), also from 2006 to 2007 surveys (Fig. 1, Table 1). Additional archived tissue samples of eight *Sebastes norvegicus* and seven *Sebastes viviparus* were also collected on board the Irish research vessel *RV Celtic Explorer* in May 2012 off Newfoundland, Canada. The sampling scheme and sample sizes per site are summarized in Table 1.

**Generation of molecular data**

DNA was extracted using a modified salt extraction protocol (Miller et al. 1988). An approximately 500-base pair (bp) fragment of the non-coding mitochondrial control region was amplified using primers developed by Hyde & Vetter (2007; D-REF: 5′- CCT GAA AAT AGG AAC CAA ATG CCA G-3′ and Thr-REF: 5′- GAG GAY AAA GCA CTT GAA TGA GC-3′). The intron-free rhodopsin gene was used to amplify an approximately 800-bp fragment in *S. mentella* samples using primers developed by Chen et al. (2003): Rh193, 5′- CTG ATG AAT AYC CTC AGT ACT ACC-3′ and Rh1039r, 5′- TGC TTG TTC ATG CAG ATG TAG A-3′. PCRs and temperature profiles followed Shum et al. (2014). PCR products were purified and sequenced by Source BioScience LifeSciences (http://www.lifesciences.sourcebioscience.com/). Resulting sequences were manually checked for quality, edited and trimmed using CHROMAS LITE v2.1.1 (http://technelysium.com.au/?page_id=13) and aligned using MUSCLE v3.7 (Edgar 2004).

**Data analysis**

**Mitochondrial DNA variation and structure.** Molecular diversity indices, including nucleotide (π) (Nei 1987) and haplotype (θ) (Nei & Tajima 1981) diversities, were estimated using DNAsp v5.10 (Librado & Rozas 2009). Haplotype genealogies were constructed in the program HapView, following a method described by Salzburger et al. (2011), based on a maximum-likelihood tree implemented in PHYLIP v3.695 (Felsenstein 1989) for mtDNA. To gauge the level of population differentiation among collections, ARLEQUIN v.3.5.1.2 (Excoffier & Lischer 2010) was used to calculate pairwise population ΦST and FST, with significance of pairwise differences at the level of 0.05 assessed with 10 000 permutations. As a correction for multiple tests, P-values were adjusted according to the modified false discovery rate method (Narum 2006). The relationship among all sample collections was visualized by multidimensional scaling (MDS) of pairwise ΦST and FST using the MASS (Venables & Ripley 2002) package in the R programming environment (Team RDC, 2005). Spatial analysis of molecular variance (SAMOVA 1.0) (Dupanloup et al. 2002) was used to identify distinct groups of populations within which (FSC) genetic variance is minimal and among which (FCT) it is greatest. This method partitions the populations into a specified number of groups, and the partition scheme (k) that maximizes differences between FSC and FCT is selected. The optimum number of groups was determined by running SAMOVA with two to eight groups with 100 annealing steps for each run.

**Distribution of geographically restricted alleles SAShA** – We tested the extent to which haplotypes are randomly distributed in the NA, as implemented in the program SAShA (Kelly et al. 2010). Assuming haplotypes are identical by descent, nonrandom distribution of haplotypes can indicate departures from panmixia and occurrences of the same haplotypes in different locations can be considered evidence of gene flow. SAShA generates observed distribution (OM) of geographic distances among instances of each haplotype which is compared to the null distribution (EM; i.e. allele distribution under panmixia) generated from the same data. An OM significantly less than EM indicates that haplotypes are underdistributed and that gene flow is somewhat restricted. For mtDNA, we tested for significant deviation of the arithmetic mean of OM from EM (Dg) using 1000 nonparametric permutations of the haplotype-by-location and haplotype-by-depth data set.

**Demographic changes.** First, Tajima’s DT (Tajima 1983) and Fu’s Fs (Fu 1997), calculated in DnaSP, were tested to examine deviations of the mitochondrial site frequency spectrum expected under the neutral expansion model for major clades (see Results). Significant and negative D T and Fs values indicate population size changes or directional selection (i.e. selective sweep; Aris-Brosou & Excoffier 1996). Mismatch distribution

© 2015 The Authors. *Molecular Ecology* Published by & Sons Ltd.
analysis was also carried out, as implemented in ABLE-QUIN, whereby a unimodal frequency distribution of pairwise haplotype differences is expected for populations that have experienced recent demographic expansion, and a multimodal frequency distribution is expected for populations at equilibrium. To estimate the divergence time, we used the formula $T = Da/2\mu$, where $2\mu$ represents a general mtDNA evolutionary rate, commonly assumed to be around 11% per million years for fish mtDNA control region (Patarnello et al. 2007).

To further assess the demographic changes in effective population size since the time of most recent common ancestor (TMRCA) of the identified lineages, Bayesian skyline plots (BSPs) were generated using the software package BEAST v. 1.8 (Drummond et al. 2012). Analysis was performed using the best fit model of nucleotide substitution (GTR+$\Gamma$) implemented using MODELTEST3.7 (Hasegawa et al. 1985; Posada & Crandall 1998), and a fixed clock was set using 11%/million years (Myr) (Patarnello et al. 2007). For each analysis, the Monte Carlo Markov chain (MCMC) was set at 150 million steps, which yielded effective sample sizes (ESS) of at least 200. Once the appropriate mixing and convergence was met, the first 10% of the posterior was discarded and the remainder combined for parameter inferences. BSPs were estimated in Tracer v. 1.6 (Rambaut et al. 2014) and plotted using the upper 95% highest posterior density.

**Marker comparison and approximate Bayesian computation.** To gain an exhaustive picture of genetic variation, we also investigated the nuclear intron-free, gene coding for the rhodopsin pigment, which Shum et al. (2014) reported to also exhibit alternative genotypes associated with depth. Haplotype genealogies for 160 _S. mentella_ rhodopsin sequences were constructed in HapView using a maximum-likelihood tree generated using PHYLIP. MtDNA haplogroup distributions and rhodopsin genotypic frequencies for the shallow- and deep-caught _S. mentella_ groups were compared using $\chi^2$ tests ($2 \times 2$ contingency tables). Furthermore, we re-examined microsatellite data at 12 loci, previously genotyped by Stefánsson et al. (2009a), for the 118 historical samples from 2006 to 2007 (Stefánsson et al. 2009a), and reanalysed them in combination with the newly generated mtDNA sequences. Population structure was assessed by calculating pairwise genetic differentiation through Slatkin’s $R_{ST}$ and Weir & Cockerham’s $F_{ST}$, with 9999 permutations carried out to obtain significance levels, using GenAIEx 6.501 (Peakall & Smouse 2006). $F_{ST}$ measures genetic differentiation based on allele identity, whereas its analogue $R_{ST}$ is an allele size measure of differentiation that assumes a strictly stepwise-mutation process. $R_{ST}$ is expected to be larger than $F_{ST}$ if populations have diverged for a sufficiently long time in the case of ancient isolation (Hardy et al. 2003). To visualize the relationship among samples, MDS analysis based on $F_{ST}$ and $R_{ST}$ values was carried out in the r environment, and population structure of individual genotypes was visualized by correspondence analysis (CA) using GENETIX 4.05 (Belkhir et al. 1996).

We tested concordance of pairwise genetic distances using a simple Mantel test among redfish collections from available areas, based on mtDNA $\Phi_{ST}/F_{ST}$ vs. microsatellite $F_{ST}$. To further illustrate the distribution pattern of horizontal and vertical genetic variation, partial Mantel tests were calculated between matrices of genetic distances (using mtDNA and microsatellite markers) and depth (m) while removing the influence of geographic distance (km), and vice versa, using the VEGAN (Oksanen et al. 2011) package in R, with statistical significance evaluated with 9999 permutations.

We then used approximate Bayesian computation (ABC), implemented in DIYABC v2.0 (Cornuet et al. 2008, 2014), to assess the evolutionary relationships among _S. mentella_ lineages in the North Atlantic. We compared six simple scenarios (see Results), to identify the most likely evolutionary events, and the effective population sizes and times of divergence implicated. The ABC approach is used to provide inference about the posterior distribution of the model parameters in order to establish how the summary statistics compare between simulated and observed data sets (Cornuet et al. 2008). We used default priors and generated simulated data sets combining both mtDNA and microsatellite information, with a total of 28 summary statistics, the calculation of which is implemented in DIYABC; for microsatellites: mean number of alleles, genic diversity, size variance and Garza & Williamson’s M-ratio per population and $F_{ST}$ pairwise divergence; for mtDNA: $F_{ST}$ pairwise divergence only. Additional summary statistics generated for mtDNA failed to provide satisfactory statistical fit between the observed and simulated data sets (data not shown). Runs consisted of six million simulated data sets (1 000 000 million for each scenario) and were evaluated by principal components analysis (PCA), based on the obtained summary statistics of a subset of simulations, to check the threshold distance of parameter estimates between simulated and observed data. The relative posterior probability was then estimated over each scenario via a logistic regression on 1% of simulated data sets closest to the observed data set. The posterior distribution of parameters for the summary statistics associated with the retained data set was then estimated through locally weighted linear regression.
Results

Mitochondrial DNA lineages

MtDNA sequence data of 261 *Sebastes mentella* individuals from 16 collections stretching across the North Atlantic produced a 444-bp fragment alignment with a total of 44 polymorphic sites, 25 of which were parsimony informative (Accession nos: KP988027–KP988288). These polymorphisms defined 56 haplotypes with overall haplotype diversity and mean nucleotide diversity of $h = 0.897 \pm 0.009$ and $\pi = 0.005 \pm 0.0002$, respectively (Table 1). The haplotypes were organized into two main divergent haplogroups differentiated by a mean net sequence divergence percentage ($D_a$) of 2.45 ± 0.47 (Fig. 2a). These haplogroups ‘clade A’ and ‘clade B’ correspond to the ‘shallow’ and ‘deep’ groups previously described by Shum et al. (2014; and Fig. S1, Supporting information).

Among all samples, 91.7% of redfish caught above 500 m (59.5%) were classified as clade A and 8.3% as clade B haplotypes, whereas among redfish caught below 500 m (40.5%), 34% belonged to clade A and 66% to clade B haplotypes. Clade A included 68.3% of the total haplotypes found in 16 collections with 12 shared haplotypes (43%) and 16 singletons (57%). Clade B haplotypes (31.7%) formed a far more circumscribed starburst pattern, with haplotypes all found in eight of the 16 locations with six shared haplotypes (21.1%) and 22 singletons (79%), suggestive of a recent expansion. The clades generally show an association with both location ($\chi^2 = 180.75$, $P < 0.0001$, df = 15) and depth ($\chi^2 = 61.76$, $P < 0.0001$, df = 1), with only a few examples of shared haplotypes between adjacent groups (Fig. 2a). Within the Irminger Sea, clade A haplotypes dominated waters above 500 m (75%) in the northeast, west and southwest, while clade B was prevalent at depths below 500 m (90%) in the northeast and west (Fig. 1).

Overall, estimates of genetic differentiation for all comparisons involving the northeast Irminger Sea, west Faroe and Norwegian Sea samples were significant ($\Phi_{ST} = 0.055$–0.423 and $F_{ST} = 0.100$–0.635), while the remaining samples show no significant genetic structure and low heterogeneity (Table 2). The MDS plot (Fig. 3a, b) based on mtDNA ($\phi_{ST}$ & $F_{ST}$) pairwise genetic distances shows consistent subdivision of the shallow-pelagic clade A from the deep-pelagic clade B groups along axis 1. Axis 2 shows a separation between the deep-pelagic west Faroe from the NE Irminger Sea groups (>500 m).

The results of *samova* indicated significant population genetic structure for each number of $k$ groups assumed. $F_{CT}$ in the *samova* analysis showed increasing values of $k$ up to 4 and began to progressively decline after that, while $F_{SC}$ drops significantly at $k = 4$, which resulted in the maximum variance between the two indices at this point. Further increases in the number of $k$ led to a dissolution in group structure, where groups with the larger proportion of private haplotypes were singled out. Thus, we chose four subpopulations as the most parsimonious partition scheme (Fig. 4). At $k = 2$, *samova* recovered a deep mitochondrial split between the combined ‘deep-pelagic’ groups from northeast Irminger Sea (>500 m) and west Faroe Island and everything else. At $k = 3$, the northeast Irminger Sea (>500 m) group and the west Faroe group became separated. At $k = 4$, redfish from the majority of clade A caught above 500 m depth formed subpopulation SI (see Table 3), the northeast Irminger Sea and west Faroe groups (deeper than 500 m), representing the core of clade B, formed subpopulation SII and SIII, respectively; finally, some redfish from the northeast Irminger Sea (<500 m; $n = 11$, clade A: 45%, clade B: 55%) formed subpopulation SIV.

Fig. 2 Haplotype genealogy of North Atlantic *Sebastes* sequences yielded using HaplotypeViewer, using maximum likelihood. (a) 68 mitochondrial control region *Sebastes mentella* haplotypes are ordered into clade A and clade B (See Table 3); (b) three rhodopsin haplotypes ordered as shallow and deep groups. Haplotypes are coloured according to *samova* groupings, and the size of each circle is proportional to the frequency of haplotypes. The lengths of the connecting lines reflect the number of mutations between them.
Table 2. mtDNA estimates of pairwise genetic differentiation among 16 *Sebastes mentella* collections

<table>
<thead>
<tr>
<th>Region</th>
<th>SW13</th>
<th>W13</th>
<th>W06</th>
<th>NE13</th>
<th>NE06</th>
<th>NS</th>
<th>NIW</th>
<th>FW1</th>
<th>FW2</th>
<th>FE1</th>
<th>FE2</th>
<th>NS07</th>
<th>NIW06</th>
<th>FW2007</th>
<th>NE07</th>
</tr>
</thead>
<tbody>
<tr>
<td>SW13</td>
<td></td>
<td></td>
<td></td>
<td>0.033</td>
<td>0.003</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>W13</td>
<td></td>
<td>-0.20</td>
<td>-0.09</td>
<td>-0.33</td>
<td>-0.03</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>W06</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>-0.00</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>NE13</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>NE06</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>NS</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>NIW</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>FW1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>FW2</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>FE1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>FE2</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>NS07</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>NIW06</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>FW2007</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>NE07</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

ΦST below diagonal and FST above diagonal at 0.05 P-value significance following false discovery rate (FDR) correction.

ΦST values between the nine geographically defined regions ranged from -0.003 to 0.195.

A hierarchical AMOVA showed the molecular variance attributed to variance among groups to account for 37.54% (ΦCT = 0.375, *P* < 0.001), while differentiation between collections within the same group (ΦSC) was found not to be significant (ΦSC = 0.013, *P* = 0.123).

The analysis using SASHA revealed the average distance between co-occurring haplotypes was 951.9 km (geographic distance) and 178.3 m (depth), with evidence that haplotypes were geographically restricted as the observed distribution (OM) was significantly different from the expected (EM) under the assumption of panmixia for geographic distance (D* = 80, *P* = 0.018) and depth (D* = 11.6, *P* = 0.039).

Demographic analysis on a significantly expanded data set showed similar results to Shum et al. (2014): unimodal distributions with negative and significant Tajima’s D and Fu’s F values for clade A (*D* = −1.67, *P* < 0.001; *F* = −20.57, *P* < 0.001) and clade B (*D* = −2.14, *P* < 0.001; *F* = −27.09, *P* < 0.001), providing support for recent population expansion (Fig. S2, Supporting information). We find that the ‘deep’ and ‘shallow’ lineages split around 22,000 years ago. The coalescent-based Bayesian skyline plot (BSP) provided details on how mtDNA diversity changed through time, indicating a rapid population growth for clade A in the past 10–12 kyr before present (BP), with a very recent stall/reduction in effective population size (Fig. 5a). Clade B haplotypes demonstrate a signal of a sustained period of population increase dating back over 15–25 kry BP (Fig. 5b).

Comparative nuclear and mitochondrial data

Rhodopsin sequence data from 160 *S. mentella* individuals (Accession nos: KR818563–KR818700) produced a 722-bp fragment alignment with a total of two polymorphic sites at positions 208 and 228. These polymorphic sites defined three haplotypes structured into well-defined shallow and deep rhodopsin clades (Fig. 2b). The single nucleotide polymorphism (SNP) T228 was only variable among the ‘shallow’ group; however, one SNP found at position 208 was fixed for alternative genotypes being assigned to mtDNA clade B.

Microsatellite pairwise *F*ST values between the nine geographically defined regions ranged from −0.003 to 0.375.
Fig. 3 Multidimensional scaling plot (MDS) of *Sebastes* mtDNA variation from 16 collections (a) $F_{ST}$, (b) $\Phi_{ST}$; microsatellite variation from nine collections (c) $F_{ST}$ and (d) $R_{ST}$ pairwise genetic distances across the North Atlantic (Location codes (see Table 1) coloured according to SAMOVA groupings); (e) correspondence analyses of *Sebastes mentella* individuals from nine locations, based on microsatellite genotypes. Labelled genotypes showed discordance with mtDNA lineage assignment (see Table S2, Supporting information for further details).
0.040 (Table S1, Supporting information) and were significant for most combinations that included the Irminger Sea (>500 m) and Norwegian international water collections, after FDR adjustments. Overall, 13 of the 36 pairwise comparisons among historical samples resulted in a significant phylogeographic signal ($R_{ST}$ significantly larger than $F_{ST}$, see Table S1, Supporting information, Fig. 3c, d).

The correspondence analysis clearly segregated redfish into three main groups. Axis 1 separated the ‘deep-pelagic’ clade B groups (IrNEDP06 & FW1DP07/FW2DP07) from ‘shallow-pelagic’ clade A groups (IrSW06, FE1SP07/FE2SP07, NSSP07 & NIW1SP06/NIW2SP06), and Axis 2 separated the west Faroe (FW1SP07/FW2SP07) from the east Faroe (FE2SP07) and northeast Irminger Sea (>500 m: IrNE06) groups (Fig. 3e).

North Atlantic structuring of redfish based on microsatellite markers and results from mtDNA (SAMOVA) consistently divided the ‘deep-pelagic’ groups [Irminger Sea (>500 m) & west Faroe] into two subgroups. We found concordance between pairwise genetic distances among redfish in nine localities based on mitochondrial $\Phi_{ST}/F_{ST}$ vs. microsatellite $F_{ST}$, showing a significant positive correlation of the measures of genetic differentiation (Fig. S3, Supporting information, tested with 9999 permutations). We found no significant signal of isolation by distance ($r = 0.12$, $P = 0.24$) or depth ($r = 0.12$, $P = 0.25$) based on microsatellite $F_{ST}$, using a partial Mantel test where the depth/geographic distance matrix was held constant. For mtDNA, partial Mantel tests showed a significant correlation between $\phi_{CT}$ and $F_{ST}$. 

### Table 3: Analysis of molecular variance for SAMOVA groupings for Sebastes mentella from 16 localities in the North Atlantic

<table>
<thead>
<tr>
<th>Groups</th>
<th>Group names</th>
<th>Tajima’s D</th>
<th>Fu’s Fs</th>
<th>Source of variation</th>
<th>Percentage of variation</th>
<th>Fixation indices</th>
</tr>
</thead>
<tbody>
<tr>
<td>IrWEP06 + IrWDP13 +</td>
<td>SI $-1.82^{**}$</td>
<td>$-22.62^{***}$</td>
<td>Among groups</td>
<td>37.54</td>
<td>$F_{CT} = 0.375^{***}$</td>
<td></td>
</tr>
<tr>
<td>IrSW06 + IrSWDP13 +</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>IrSWEP06 + FE1SP07 +</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>IrSWDP06 + FE2SP07 +</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>IrSW06NIW2SP06</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>IrNE06 + IrNEEP13</td>
<td>SI $-2.02^{**}$</td>
<td>$-16.45^{***}$</td>
<td>Among populations</td>
<td>0.84</td>
<td>$F_{SC} = 0.013^{***}$</td>
<td></td>
</tr>
<tr>
<td>FW1SP07 + FW2SP07</td>
<td>SIII $-0.47$</td>
<td>$-5.17^{**}$</td>
<td>Within populations</td>
<td>61.62</td>
<td>$F_{ST} = 0.383^{***}$</td>
<td></td>
</tr>
<tr>
<td>IrNEEP13</td>
<td>SIV 1.43</td>
<td>2.47</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

<0.05 Bold; **<0.01; ***<0.001

© 2015 The Authors. Molecular Ecology Published by & Sons Ltd.
genetic distance ($F_{ST}$) and depth ($r = 0.329$, $P = 0.019$), but not for geographic distance ($r = 0.009$, $P = 0.407$).

Given the detection of geographically disjunct ‘deep’ populations in both the Irminger and Faroese seas, we tested six simple historical demographic scenarios involving pairs of ‘shallow’ and ‘deep’ populations from these areas (bearing in mind the relatively consistent homogeneity on the ‘shallow’ populations all across the NA, Figs 3 and 6). The first scenario involved an original split between the ‘deep’ and ‘shallow’ lineages followed by subsequent split between the Irminger and Faroe seas within both deep and shallow lineages, leading to the current geographical disjunction. The second scenario involved a parallel, independent origin of deep sea groups from their shallow ancestors, respectively, in the Irminger and Faroese seas. The third differed from the second scenario with an independent origin of the shallow groups from their deep sea ancestors. The fourth, fifth and sixth scenarios involved a simplistic tree-like bifurcations occurring from time ta to t1 in the past. Estimations of the posterior probability based on both direct and logistic regression for each scenario provided unambiguous support for the scenario S1, with a probability of 91% and a 95% CI of 90–92, not overlapping with any other scenarios (Fig. 6). Given a generation time between 12 and 20 years (Stransky et al. 2005a,b), the scenario assumes that the ‘deep–pelagic’ and ‘shallow–pelagic’ redfish split ~4500–7500 years ago (ta median = 374 generations, 95% CI: 93.4–419). The subsequent split between the ‘deep–pelagic’ Irminger Sea (IrNE06: $N_e = 8570$, 95% CI: 5840–9900) and west Faroe (FW06: $N_e = 7040$, 95% CI: 3160–9800) shares similar divergence times as the split between the ‘shallow–pelagic’ Irminger Sea (IrSW06: $N_e = 6610$, 95% CI: 3310–9640) and east Faroe (FE06: $N_e = 5570$, 95% CI: 2020–9510), estimated at approximately ~1200–2000 years ago (t1 median = 98.3 generations, 95% CI: 27.9–281).

Discussion

Based on greatly expanded geographical screening, sample sizes and analytical toolkit, this study lends strong support to the existence of at least two highly distinct evolutionary units of oceanic Sebastes mentella in the North Atlantic (NA). Nuclear patterns are mirrored and strengthened by mitochondrial evidence, and, perhaps more strikingly, the pattern of divergence is strongly associated with habitat depth (more so than with geographical distance). Coalescent patterns suggest that the emergence of depth-related structure may have originated in the Late Pleistocene, with the two lineages segregating after the recolonization of the Irminger Sea, as the ice sheets retreated at the end of the last glaciation.

Population structure and connectivity

Analyses of mtDNA sequences revealed substantial population structure among depth-defined habitats between two distinct S. mentella mtDNA clades (A: above and B: below 500 m; Figs 1 and 2). Our results expand on recent findings, indicating that the ‘shallow’ and ‘deep’ groups in Shum et al. (2014) correspond to clades A and B identified in this study (Fig. 5), Supporting information). We found nonrandom association of haplotypes in the NA, with clade A commonly found above 500 m southwest and west of the Irminger Sea, east of the Faroes, and Norwegian waters, whereas clade B haplotypes are more localized to the central NA, below 500 m, northeast of the Irminger Sea and west of the Faroes (Fig. 1). The overall pattern of mtDNA variation is associated with isolation by depth, mirrored by rhodopsin SNP distribution, similarly detected using microsatellites (Stefansson et al. 2009a) and to some degree also supported by subtle phenotypic variation (Magnússon & Magnússon 1995; Stransky 2005; Stefansson et al. 2009a,b; Cadrin et al. 2010). MtDNA and nuclear data consistently identified the main genetic partitioning – shallow vs. deep – as well as detecting additional population subdivision, between the ‘deep–pelagic’ Irminger Sea and west Faroe groups. According to the SAMOVA and MDS analysis, redfish was further divided into four genetically distinct subclades: (SI) southwest and west Irminger Sea/east Faroe/Norwegian shelf and Norwegian international waters; (SII) northeast Irminger Sea (>500 m); (SIII) west Faroe; and (IV) northeast Irminger Sea (<500 m). However, it is worth mentioning the potential issue of small sample size for some collections as is the case of the final SAMOVA group (SIV), which probably represents an area of contact between haplogroups and may not reflect a ‘true’ distinct biological unit but a mere artefact due to low sample size. Support for this group structure is mirrored by close inspection at nuclear markers. The rhodopsin SNP shows a strong association with the mtDNA clade A and clade B haplotypes, suggesting that evolutionary independence between shallow and deep lineages is unambiguous. The two deep-pelagic groups were not statistically distinguishable at microsatellite loci, even using greater sample sizes (Stefansson et al. 2009a). Yet, analysed at mtDNA, they showed significant genetic structure (Figs 2a and 3). Strikingly, while the average mtDNA-based $F_{ST}$ variance among ‘shallow’ aggregations is nonsignificant ($F_{ST} = 0.020$, $P = 0.074$), suggesting significant large-scale connectivity, the two ‘deep’ groups, in the Irminger Sea and western Faroes, appear significantly differentiated ($F_{ST} = 0.176$, $P > 0.001$), with only three haplotypes shared, indicating that habitat segregation may have
shaped diverging behaviours and life history adaptations in these two lineages. Such a segregation of the mitochondrial matrilinage, between deep Irminger and Faroese fish, alongside a more blurred boundary showed by microsatellites, raises the possibility that female *S. mentella* in the deeper sea layer may exhibit some degree of female philopatry/residency (Petit & Excoffier 2009). The combined use of mitochondrial and nuclear information on a few selected individuals can also shed some further light on the biology and behaviour of these organisms. For example, one fish caught above 500 m from east of the Faroes (FE1SP07_8.9) exhibits a clade B haplotype, typical of the ‘deep’ layer, and its microsatellite genotype falls within the ‘deep’ group. This highlights the ability of redfish to exhibit a degree of pelagic–demersal mixing between clades from shallow and deep environments (Planque et al. 2013). Most notably, four individuals from the 2013 Irminger

![Diagram](image-url)
Sea ‘deep’ sampling (I.D. 17, 30, 37 and 40) exhibited fully shallow mtDNA and rhodopsin types (Table S2, Supporting information), which suggests that ‘shallow-to-deep’ contemporary forays might be more common than those in the opposite direction. The inception of this behaviour may originate during the establishment of juvenile ‘site fidelity’ during the early stages of red-fish development. In the Irminger and Norwegian seas, spawning female redfish are distributed along the larval extrusion areas of the Reykjanes Ridge and the Norwegian continental shelf between March and May (Magnússon & Magnússon 1995; Drevetnyak & Nedreaas 2009). The pelagic-fry drift to nursery areas found along the coast of east/west of Greenland and the Barents Sea where juveniles settle to the bottom until they mature and migrate to distributed populations throughout the North Atlantic. Yet, the complex behaviour, areas of copulation and seasonal migration patterns of this species across shallow- and deep-pelagic habitats remain poorly understood (Planque et al. 2013). Hence, this pattern suggests the occurrence of short-time migration from deep-pelagic individuals, supporting the notion of vagrants as individuals move to shallow and deeper waters during their life cycle (Shum et al. 2014).

Several examples from the east/west Faroe groups caught in both shallow and deep layers display discordant molecular markers: two samples (FE1sp07_8.5 & FE1sp07_8.6) caught approximately at 400 m display ‘deep’ haplotypes but a microsatellite genotype typical of the shallow layer. Similarly, five samples caught in deep waters (FW1dp07_7.6, FW1dp07_7.8, FW2dp07_7.15, FW2dp07_7.21 & IrNEdp06_29) exhibited clade A haplotypes but possess multilocus microsatellite genotypes falling within the ‘deep’ group. The nuclear-coding rhodopsin gene is a powerful tool in the identification of closely related fish species (Rehbein 2013; Pampoulie et al. 2015), and here, we found that only 16 of 36 specimens (4%) analysed at both mtDNA and rhodopsin yielded ambiguous assignment to their lineage of origin (Table S2, Supporting information). Interestingly, two of these individuals possessed double peaks, or heterozygous indels, in the rhodopsin chromatograms (Fig. S4, Supporting information; Sousa-Santos et al. 2005), which, along with the observed mtDNA/microsatellite mismatches, is best explained by the occasional occurrence of introgressive hybridization between the two genetically distinguishable groups. Roques et al. (2002) indicated significant introgressive hybridization between S. mentella and Sebastes fasciatus in the Gulf of St. Lawrence and surrounding waters; Pampoulie & Danielsdottir (2008) also detected signatures of hybridization between S. mentella and Sebastes norvegicus. Thus, it is reasonable to expect that infraspecific mating between a deep-pelagic clade B male and shallow-pelagic clade A females (and vice versa) would have produced the genotypic ambiguities mentioned above. The shallow- and deep-pelagic groups appear to occupy their preferential depth range in near sympathy. However, spatial overlap is more apparent to the west and northeast of the Irminger Sea and in the Faroese waters; thus, the opportunity for introgressive hybridization may derive from hybrid zones as a result of secondary contact where S. mentella are within their preferred geographic and vertical cruising limits. Pampoulie & Danielsdöttir (2008) have reported significant levels of hybridization in the NA among redfish which may indicate that different S. mentella lineages were allopatic before secondarily coming into contact to form their current sympatric distribution (Cadrin et al. 2010). Despite the potential for interbreeding, however, the shallow and deep types maintain a striking overall integrity, which bears resemblance to some notable cases of parapatric speciation (Allender et al. 2003; Berner et al. 2009; Nosil 2012).

**Historical reconstruction**

The glaciation and interglacial periods of the Pleistocene have been known to considerably shape the current distribution and connectivity among contemporary populations (Bigg et al. 2008; Finnegan et al. 2013). Evidence from mtDNA haplotypes alone suggests that the shallow- and deep-pelagic clades diverged over 22 000 years ago, in correspondence with the last glacial maximum (LGM). The Bayesian skyline plot (BSP) provides information on how mtDNA diversity changed through time, back to the most recent common ancestor. The BSP analysis indicate a gradual signature of expansion 15 000–25 000 years ago for the deep-pelagic clade, and a greater, steady increase in population growth – compared to the shallow-pelagic clade – which shows a much sharper rapid expansion around 10 000–15 000 years ago. The BSPs show contrasting signatures of population growth suggestive of postglacial influence. The expanding sea ice may have driven S. mentella to southerly latitudes as far as the Grand Banks to deeper waters during the LGM, as global sea level fell by 120–135 m and intense calving in the northern hemisphere ice sheets 18–15 kyr BP resulted in massive icebergs advancing into the North Atlantic 40°N (Grousset et al. 1993). As the sea ice retreated, characterized by intense warming and rising sea levels 14 600–13 800 years BP, S. mentella had the opportunity to advance in a northerly direction to glacial refugia south of Iceland before sea ice readvanced during the Younger Dryas (YD, 13–11kyrBP) which forced the Northern Hemisphere into near-glacial conditions (Crucifix & Berger 2002). The onset of the YD
may have favoured allopatric conditions between the two clades as the deep-pelagic group occupied deeper refuges, while the shallow-pelagic rapidly spread with the retreating sea ice and rising sea level following the YD into the Norwegian and Barents seas and the Irminger Sea ($R_{ST} > F_{ST}$ Table S1, Supporting information). The microsatellite $R_{ST}$ values provide insights on the relative divergence of $S. mentella$. The eastern NA collections (FE$_{SP07}$, NS$_{SP06}$ & NIW$_{SP06}$) present a significant phylogeographic signal with respect to the Irminger Sea collections (IrSW$_{SP06}$ & IrNE$_{SP06}$), suggesting that postglacial range shifts and secondary contact events have played a significant role in shaping $S. mentella$ spatial structure.

Overall, SAMOVA, MDS and CA analyses revealed the shallow-pelagic group shows strong homogeneity across the NA, whereas the deep-pelagic groups form rather distinct ‘pockets’, suggesting their evolutionary distinction also involves different life histories, as shallow-pelagic redfish appear more prone to connectivity, whereas the deep-pelagic redfish appear to be more strictly associated with their local habitats, probably migrating less. We tested which demographic scenario could best explain the genetic patterns among pelagic pairs of shallow and deep populations in the Irminger Sea and west of the Faroes, using an ABC approach. Estimates for the posterior probability for each scenario provided robust support for scenario 1 (Fig. 6). This indicates that the shallow- and deep-pelagic gene pools split approximately 5–7 kyr BP, after the postglacial recolonization of the Irminger Sea. Furthermore, a subsequent split between the deep-pelagic groups emerged approximately 2 kyr BP. Their distributions are found within close proximity of bathymetric features (see Fig. 1) along the west of the Reykjanes Ridge and the Faroe-Shetland Ridge known for complex oceanic conditions shaped by bathymetric features. Population independence was largely upheld by both nuclear and mitochondrial markers, and it is likely that depth-associated adaptive processes are at play (Shum et al. 2014) to counteract the homogenizing effects of gene flow. Given the paucity of well-characterized marine biological systems undergoing diversification processes consistent with speciation, North Atlantic $S. mentella$ should now represent a valuable subject to investigate genomic correlates and mechanisms for the maintenance of bathymetry-associated lineage sorting and, potentially, speciation.

From a practical standpoint, the present results will have cascading impacts on the assessment and management of these commercially valuable fish stocks, which remain hotly debated. Our results are consistent with two genetically distinguishable putative groups separated by depth and following different evolutionary trajectories: this should form the basis to recognize them as distinct evolutionarily significant units. Given the circumscribed local distribution of the ‘deep’ populations in the Irminger and western Faroese seas, tailored management appears required, if we are to avert the permanent loss of a unique biodiversity component.

Conclusions

Our study provided the first extensive depth-associated sampling and phylogeographic reconstruction of the North Atlantic oceanic beaked redfish, $S. mentella$. This species shows a consistent and significant distinction of at least two evolutionary lineages: one widely distributed ‘shallow’ type, which partially overlaps with local populations of a more sedentary ‘deep’ type. While evidence exists for localized interbreeding between these two lineages, the rate does not appear notably greater than similar introgressive processes still occurring among the four $S. mentella$ species in the North Atlantic. We find that mtDNA reflects redfish evolutionary history throughout the Late Pleistocene, whereas the integration with microsatellite data serves to better reflect postglacial divergence and the patterns of contemporary gene flow among populations, including the potential detection of sex-biased dispersal. The shallow-pelagic clade shows strong homogeneity across the NA, while the deep-pelagic clade is restricted to the central North Atlantic, segregated by complex oceanic conditions shaped by bathymetric features. Population independence was largely upheld by both nuclear and mitochondrial markers, and it is likely that depth-associated adaptive processes are at play (Shum et al. 2014) to counteract the homogenizing effects of gene flow. Given the paucity of well-characterized marine biological systems undergoing diversification processes consistent with speciation, North Atlantic $S. mentella$ should now represent a valuable subject to investigate genomic correlates and mechanisms for the maintenance of bathymetry-associated lineage sorting and, potentially, speciation.

Acknowledgements

We would like to acknowledge the following who contributed in the collection of biological samples: the crew and scientists on board the Icelandic t/s Ární Fríðriksson for the collection of $S. mentella$ during the redfish stock assessment survey in 2013; Killey Best for $S. fasciatus$ samples collected on board the RV Celtic Explorer by the staff and students of CFER (Center for Fisheries Ecosystems Research) during the offshore research survey in May 2012; Kjell Nedreas, Porstein Sigurðsson and Jákup Reinert for the collection of $S. mentella$ samples from Norway and the Faroe Islands. We are indebted to Valérie Chosson for her skilled technical and logistic support throughout the 2013 sampling campaign. We also thank Andrew Griffiths, Sara Vandamme, Christoph Stransky and three anonymous reviewers for their useful comments on this manuscript. This work was funded by the Marine Research Institute in Iceland and the University of Salford GTS/PtE programme.

© 2015 The Authors. Molecular Ecology Published by & Sons Ltd.
References


the role of bathymetry in the speciation process. *Molecular Ecology*, 17, 1122–1136.


S.M. and C.P. conceived the study, with contribution from P.S.; C.P. and K.K. organized the sampling, which was carried out by P.S. and K.K.; P.S. conducted all laboratory work and statistical analyses and drafted the manuscript. S.M. contributed to writing and advised on statistics, with feedback from C.P. and K.K. All authors share interests in the mechanisms of population divergence in open marine habitats and their implications for biodiversity conservation and resource management.

Data accessibility

Sequence data for this study are available via GenBank (Accession nos mtDNA: KP988027–KP988288; rhodopsin KR818563–KR818700). Microsatellite genotypes, mitochondrial DNA and rhodopsin sequences are also available via Dryad (http://dx.doi.org/10.5061/dryad.v80q9).

Supporting information

Additional supporting information may be found in the online version of this article.

Table S1 Microsatellite estimates of pairwise genetic differentiation among nine S. mentella collections.

Table S2 Genetic marker discordance among S. mentella samples for mitochondrial control region (mtDNA clades), rhodopsin SNPs and microsatellite genotype.

Table S3 Pairwise FST genetic differentiation among nine S. mentella collections. mtDNA (below diagonal) and microsatellite (above diagonal) based on 10 000 permutations.

Fig. S1 Haplotype genealogies indicating shallow- (red) and deep-pelagic (blue) samples from Shum et al. (2014) within the larger mtDNA dataset generated in the present study (See Fig. 2, main article).

Fig. S2 Mismatch distributions for oceanic S. mentella clade A (above) and clade B (below).

Fig. S3 Correlation plots between pairwise genetic distances for S. mentella collections from (a) 16 sampling regions based on mtDNA (ST vs. FST) and (b) nine sampling regions based on microsatellite and mtDNA FST. Sampling information is provided in Table 1 (main article).

Fig. S4 Chromatograms showing heterozygous SNP at position 208 of the Rhodopsin gene for (a) IrW_mar13_12 and (b) IrW_mar13_35.

© 2015 The Authors. Molecular Ecology Published by & Sons Ltd.