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Ahanchian, N, Nester, CJ, Howard, D, Ren, Lei and Parker, DJ

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Estimating the Material Properties of Heel Pad Sub-layers
Using Inverse Finite Element Analysis

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Abstract

Detailed information about the biomechanical behaviour of plantar heel pad tissue contributes to our understanding of load transfer when the foot impacts the ground. The objective of this work was to obtain the hyperelastic and viscoelastic material properties of heel pad sub-layers (skin, micro-chamber and macro-chamber layers) in-vivo.

An anatomically detailed 3D Finite Element model of the human heel was used to derive the sub-layer material properties. A combined ultrasound imaging and motorised platform system was used to compress heel pad and to create input data for the Finite Element model. The force-strain responses of the heel pad and its sub-layers under slow compression (5mm/s) and rapid loading-hold-unloading cycles (225mm/s), were measured and hyperelastic and viscoelastic properties of the three heel pad sub-layers were estimated by the model.

The loaded (under ~315N) thickness of the heel pad was measured from MR images and used for hyperelastic model validation. The capability of the model to predict peak plantar pressure was used for further validation. Experimental responses of the heel pad under different dynamic loading scenarios (loading-hold-unloading cycles at 141mm/s and sinusoidal loading with maximum velocity of 300mm/s) were used to validate the viscoelastic model.

Good agreement was achieved between the predicted and experimental results for both hyperelastic (<6.4% unloaded thickness, 4.4% maximum peak plantar pressure) and viscoelastic (Root Mean Square errors for loading and unloading periods <14.7%, 5.8% maximum force) simulations. This paper provides the first definition of material properties for heel pad sub-layers by using in-vivo experimental force-strain data and an anatomically detailed 3D Finite Element model of the heel.

1. Introduction

The behaviour of the plantar heel pad has been the topic of considerable research because it forms a critical interface with the supporting surface. It is affected by aging and disease and is the site of pain [1, 2, 3]. Study of heel pad behaviour has been achieved through experimental [4, 5, 6] and numerical methods, particularly Finite Element Analysis (FEA) [7, 8, 9]. The latter provides data such as the
distribution of internal tissue stress that cannot be experimentally measured. However, for FEA models to prove effective they should be based on geometric and material properties that ensure the model behaviour is sufficiently close to \textit{in-vivo} heel pad behaviour, as seen during human gait. In most Finite Element (FE) models, hyperelastic rather than viscoelastic material models were used to simulate nonlinear behavior of the heel pad \cite{7, 8, 9, 10}. Results from these studies were limited to static or fixed loading rates due to the absence of a dynamic \textit{in-vivo} system that allows compression of plantar tissues at various high speeds, whilst also providing the data required for estimation of viscoelastic parameters and validation.

In addition, the heel pad is typically modelled as a homogeneous single-layer material rather than an \textit{in-vivo} tri-layer biological structure (macro, micro and skin layers) \cite{7, 10, 11}. In a few cases, the heel pad was modelled as a dual-layer composite structure (fat and skin), but this ignores the different behaviours and interactions between micro and macro layers \cite{8, 9, 12}. This may compromise the ability of FEA to predict internal stresses. A further issue with some of the models reported thus far is that experimental data were obtained \textit{ex-vivo} \cite{12, 13, 14, 15}. Tissue dissection disrupts the normal \textit{in-vivo} tissue constraints and the effects of time and loss of vascular supply are not fully understood \cite{16}. Clearly, \textit{in-vivo} methods at appropriate loading rates are preferred over \textit{ex-vivo} approaches.

In summary, most of heel pad models are limited by excluding viscoelastic effects and/or using less than three layers. Moreover, some approaches to validation may not test models with sufficient rigour. Hence, the objective of this work was to estimate hyperelastic and viscoelastic material properties of ‘macro-chamber’, ‘micro-chamber’ and ‘skin’ layers using inverse FEA and \textit{in-vivo} experimental data.

2. Methods

2.1 Finite Element Model
An anatomically detailed model of the right heel of a healthy female volunteer (34 years old, height 164cm, weight 63kg, shoe size 5UK) was constructed based on unloaded MRI images. MRI data were T1 weighted with a flip angle of 25, taken in coronal view using 3D fast field echo (Philips 1.5T Acheiva), with pixel size=0.29mm×0.29mm (2.4% resolution), and slice intervals=1.25mm. The images were segmented to identify the plantar fascia, muscle tissue, macro-chamber, micro-chamber and skin layers and create corresponding 3D surface geometries using ScanIP v3.1 (Simpleware Ltd, Exeter, UK). Different segmentation algorithms including thresholding, confidence connected region-growing, floodfill and paint were used for identifying the corresponding tissues. 3D surface geometries were imported into SolidWorks 2010 (Dassault Systemes, USA) to generate 3D solid geometries and the complete assembly. Since MRI slices were out of the plane of boundaries between soft tissue layers, the effect on structural modelling will be minimal. Also, the 0.29mm between slices is a small percentage of the anterior/posterior length of the structured modelled. A full description of the development of the heel region structures can be found elsewhere [17].

To reduce the computation time only a portion of the foot was modelled. Planes at 92.5mm from the back and 45mm from the bottom of the heel were chosen to be flat face boundaries of the model. The solid model was meshed with 11,504 hexahedral elements (type C3D8R) using ABAQUS v6.10 (Dessault Systemes, USA). The number of elements was obtained by performing a mesh convergence study. The selected mesh density was based on the change in the peak force for a subsequent doubling of mesh density being less than 3%. The meshed model was exported to Ls-Dyna v2.2 (Livermore Software Technology Corporation, Livermore, USA) for inverse FEA. Effects of stiff tissues (foot bones and Achilles tendon) on the biomechanical behaviour of the heel pad were simulated by applying zero-displacement constraints to all nodes forming the soft tissue-stiff tissue interface. The Achilles was modelled as stiff since under tension it will be far stiffer than the fat pad and far from it too, acting as a rigid attachment to the heel bone which is thereafter attached to the heel pad. All nodes at the superior and anterior boundaries (flat faces) of the model were fully constrained. The model was tilted by 17° to replicate the position of the foot during subsequent experiments performed with a Soft Tissue Response Imaging Device (STRIDE) (Figure 1) [18]. In Ls-Dyna the flat
indentation plate of the STRIDE was modelled as a rigid structure (Figure 1). Tied contact was defined between the parts of the heel model and frictionless surface-to-surface contact was defined between the indentation plate and heel skin.

The macro-chamber, micro-chamber and skin were modelled as nonlinear viscoelastic materials (Figure 1). The first-order Ogden model was used to represent the hyperelastic behaviour of heel pad tissues as done previously [7, 8, 9]. The corresponding material properties appear in the strain energy function as follows

\[ W = \frac{\mu}{\alpha} (\lambda_1^a + \lambda_2^a + \lambda_3^a - 3) \]  

where \( \lambda_1, \lambda_2, \lambda_3 \) are the principal stretches in the x, y and z directions respectively, \( \mu \) is the shear modulus, and \( \alpha \) is the deviatoric exponent (\( \mu \) and \( \alpha \) being the hyperelastic material parameters). Viscoelastic tissue behaviour was modelled using one generalized Maxwell element for the viscoelastic overstress in the Ogden model. The Maxwell viscoelastic element consists of a linear spring with stiffness \( G_1 \) and a linear damper with viscosity \( \nu_1 \) in series. The relaxation time (a measure of the time taken for the stress to relax) for the Maxwell unit is \( \tau_1=\nu_1/G_1 \). Its inverse is the decay constant \( \beta_1=1/\tau_1 \). The stiffness \( G_1 \) (the shear relaxation modulus) and decay constant \( \beta_1 \) are the viscoelastic material parameters of the model in LS-Dyna. The corresponding material properties appear in the relaxation function, \( G(t) \), written as a first-order Prony series, representing the combined hyperelastic and viscoelastic model as follows

\[ G(t) = G_\infty + G_1 e^{-\beta_1 t} \]  

where \( G_\infty \) is the long term shear modulus (Figure 1).
The focus of the reported work is to identify the properties of and model the heel pad. However, the surrounding tissues that constrain the heel pad must also be modelled adequately enough to provide realistic boundary conditions. Therefore, to simplify the FE model, the plantar fascia and muscle tissues were modelled as linear elastic materials. However, the literature contains limited reports concerning the material properties of muscle tissues and plantar fascia, and, in most other FE studies, the foot muscles have been merged with the heel pad tissue and assigned the same material properties [19, 20, 21]. Moreover, the plantar fascia has previously been modelled with tension-only truss elements with Young’s modulus determined from tensile tests [19, 20, 22]. Since there is poor agreement between studies, a series of parametric studies was conducted to assess the sensitivity of the FEA results to the material properties used for the plantar fascia and muscle tissue. Different material properties, derived from published data [21, 22, 23, 24, 25], were assigned to the plantar fascia and muscle tissue and this revealed only a small effect on the force-strain behaviour of the heel pad (Root Mean Square (RMS) error <1.5% and <0.67% max force for the plantar fascia and muscle tissue respectively). The initial material properties derived from published literature were therefore used to start the FEA (Table 1).

### Table 1
Initial material properties of each component in the FE model

<table>
<thead>
<tr>
<th>Material model</th>
<th>Material properties</th>
<th>Poisson’s ratio</th>
<th>Density (g/mm$^3$)</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>Muscle tissue</td>
<td>Linear elastic</td>
<td>0.49</td>
<td>1×10$^{-3}$</td>
<td>[22]</td>
</tr>
<tr>
<td>Material</td>
<td>Type</td>
<td>Modulus (MPa)</td>
<td>Poisson's Ratio</td>
<td>Shear Modulus (MPa)</td>
</tr>
<tr>
<td>--------------------------</td>
<td>---------------</td>
<td>---------------</td>
<td>-----------------</td>
<td>---------------------</td>
</tr>
<tr>
<td>Plantar fascia</td>
<td>Linear elastic</td>
<td>350</td>
<td>0.40</td>
<td>$1 \times 10^{-3}$</td>
</tr>
<tr>
<td>Heel pad sub-layers</td>
<td>Hyperelastic</td>
<td>0.016</td>
<td>0.4999</td>
<td>$1 \times 10^{-3}$</td>
</tr>
<tr>
<td></td>
<td>Viscoelastic</td>
<td>0.389</td>
<td>1000</td>
<td>$8.3 \times 10^{-3}$</td>
</tr>
<tr>
<td>Indentation system</td>
<td>Rigid</td>
<td>2.07</td>
<td>0.3</td>
<td>$7.83 \times 10^{-3}$</td>
</tr>
</tbody>
</table>

2.2 Experimental Acquisition of Force and Tissue Displacement Data

The aim of this stage was to perform a series of slow and rapid compression tests on the same heel used to generate the geometric model and obtain the force-strain responses of the heel pad and its sub-layers. Ethical approval was granted by the University of Salford ethical committee.

STRIDE applies controlled vertical compression cycles of various speeds and load profiles to the heel pad in-vivo. It simultaneously uses an ultrasound system with a 5.5MHz probe in B-Mode and capture frequency of 201Hz (MyLab 70, Esaote, Italy) with a measurement accuracy of 1.75% (±0.7mm) to track changes in the heel pad and the boundaries between its sublayers during loading/unloading. STRIDE uniformly compresses the heel using a 150mm diameter flat rigid steel plate. A 20mm diameter circular plastic window at the centre of the plate allows imaging of the tissue. Example of ultrasound images (for the unloaded and loaded heel pad) is shown in Figures 2. The boundary of the calcaneus can be seen as a white, thick arc at the lower part of the ultrasound images. The interface between the macro-chamber and micro-chamber layers is the indistinct thick white layer in the middle of the ultrasound images. However, by adjusting the brightness and contrast of the images, this boundary becomes much clearer at the expense of the other features. The boundaries of the skin layer are thin white bands, one adjacent to the plastic window and the other forming the interface with the micro-chamber layer. As can be seen, static ultrasound images are difficult to interpret. However, the ultrasound videos of the indentation process clearly show the moving boundaries and then the boundaries in the corresponding static images can be identified by cross-referencing with the videos (Video 1). The ultrasound images were used to measure the unloaded and loaded thickness (UT and LT) of the heel pad, macro-chamber and micro-chamber layers in the vertical Y direction (i.e.
perpendicular to the flat indenter surface in Figure 2). The engineering strains of the three tissue layers were then calculated as follows:

$$\varepsilon = \frac{U_T - L_T}{U_T}$$  \hspace{1cm} (3)

These measurements were taken under the calcaneus tuberosity above the plastic window (Figure 2). The vertical compression force in the Y direction, F in Figure 2, applied to the tissue above the window is measured independently of the total load applied to heel area using a miniature load cell (500lb Precision, 3000Hz, TC34, Amber Instrument, UK) with linearity of 0.02% (4.45N). The load recorded under the heel pad by the miniature load cell versus strains of heel pad, macro-chamber and micro-chamber was used as input to the FEA. All tests were done while the subject was standing and the calcaneus tuberosity located above the centre of the window. The foot was in a foot brace (Aircast boot) which allowed vertical compression of the heel without lifting of the foot (Figure 2).
Figure 2. Soft Tissue Response Imaging Device (STRIDE) and ultrasound images for the frontal view at the location of calcaneus tuberosity: (A) Isometric view of STRIDE; (B) Cross-section and foot brace arrangement; (C) Unloaded heel pad; (D) Loaded heel pad

The moving boundaries of the heel pad sub-layers can be seen more clearly in the ultrasound video clip recorded during compressing of the heel pad by STRIDE (Video 1).

Ultrasound video of compression tracking.mp4

Using STRIDE, slow compression tests at 5mm/s and rapid compression tests at 225mm/s (comparable to the velocity of vertical impact in slow walking) were performed in order to determine the material properties of the heel pad sublayers. For these tests, the indenter followed a truncated triangular waveform consisting of 4 phases: load at constant speed; a 26ms hold period; unload at constant speed; a 26ms hold period. For validation of the viscoelastic FE model, another two different loading cycles were applied: (1) load/unload at a constant speed of 141mm/s (with 26ms hold), and (2) sinusoidal loading-unloading cycles with a maximum speed of 300mm/s. These achieved compression of up to 36.5% (5.7mm) the unloaded thickness of the heel pad. The compression tests were repeated for five iterations with 1-minute rest between each trial to allow for tissue recovery. The unloaded thickness of the heel pad sub-layers was measured from the first available ultrasound image i.e. when the indenter first touched the plantar tissue.

The force-strain responses of the heel pad and its sub-layers indicated that their behaviours are nonlinear with an initial low stiffness region, followed by increasing stiffness. The results showed that the macro-chamber, micro-chamber and skin layers formed 76.4, 14.7, and 8.9% of the unloaded heel pad thickness respectively. Test results showed that the resistance of the heel pad is increased by increasing loading velocity. During slow compression (5mm/s), an average load of ~73N was required to obtain a 36.5% strain of the heel pad, whereas ~96N and ~114N were required at constant velocities of 141 and 225mm/s. The increase in loading velocity resulted in an increase in Energy Dissipation Ratio (EDR). For compression at 141mm/s, EDR was 63.3%, whereas it was 76.1% at
225mm/s. Under sinusoidal loading EDR was measured as 78% that is close to results for impact and ballistic pendulum tests performed on healthy adults (79-90%) [27].

2.3 Inverse Finite Element Analysis

The inverse FEA procedure was broken into multiple stages (associated with the different tissue layers) as shown in Figure 4. This procedure was used twice: firstly to estimate the hyperelastic parameters and then to estimate the viscoelastic parameters. In this way, at each stage only two material properties had to be found, which was done using the manual search technique summarised in Figure 3. The latter will be explained first before describing the multiple stages associated with the different tissue layers.

The force–strain responses of the heel pad and its sublayers (Figure 6), obtained from the physical tests, were used as inputs to the manual searches (the FEA model itself being driven by the corresponding indenter motion profiles). Referring to Figure 3, the comparison between experiment and FEA was based on the RMS force error and the difference between maximum strains (calculated using Excel). The RMS error was calculated as follows:

\[
RMS \text{ error} = \frac{\sqrt{\sum_{k=1}^{n} (F_{ke} - F'_{ke})^2}}{n} \tag{4}
\]

where \(F_{ke}\) and \(F'_{ke}\) are model predicted and experimental forces respectively, and \(k\) is the data point index. In each manual search, the magnitudes of the adjustments made to the two material properties (e.g. \(\mu\) and \(\alpha\), for the current tissue layer, were chosen so that the FEA outputs moved gradually towards the experimental results (RMS error decreasing). When the RMS error passed a minimum and started to increase, the adjustments were halved and their sign changed. In this way, the minimum RMS error was found.
In the first stage, the macro-chamber layer FE elements (i.e., a one layer model) were used to determine first estimates of the macro-chamber material properties, which were assigned with initial values of $\mu=0.016$ MPa and $\alpha=6.82$ [7]. These properties were then adjusted using the manual search procedure described above to optimise the fit with the experimental data for the macro-chamber layer. This process was repeated for 21 iterations until no useful reduction was observed in the RMS error (i.e., when the change in RMS error between the last two iterations was less than 0.2\% of the maximum force). The parameters determined at this stage were not the final values since they were obtained in the absence of the constraints applied by the micro-chamber and skin layers in-vivo.
Inverse FEA procedure for estimating the material properties of the macro-chamber, micro-chamber and skin layers.

In the second stage, the elements representing the micro-chamber were added to the model (i.e. a two-layer model was created). Micro-chamber properties were adjusted iteratively, starting from properties derived for the macro-chamber layer, to optimise the fit with the experimental data for the combined...
macro-micro layers. Additional constraints applied to the macro-chamber layer by the micro-chamber layer inevitably affected the response of the macro-chamber layer. Therefore, the macro-chamber behaviour was reviewed during each iteration alongside the adjustment of micro-chamber material properties, its properties being varied to optimise the fit with the macro-chamber experimental data.

The process of adjusting the material parameters of the macro-chamber and micro-chamber layers was repeated for 23 iterations until the objective functions of the macro-chamber layer and two-layer model did not change significantly between iterations (i.e. when $\Delta \text{RMS} < 0.5\%$ of maximum force and $\Delta \text{maximum strain} < 0.05\%$ of maximum strain respectively).

In the final stage, the complete model incorporating macro-chamber, micro-chamber and skin layer (i.e. a three-layer model) was used for estimation of the final values of the material parameters of the heel pad sub-layers. Skin properties were adjusted in an iterative procedure, starting from properties derived for the micro-chamber layer, to optimise the fit between predicted results for the complete model and the experimental data for the entire heel pad. Additional constraints applied to the micro and macro-chamber layers by skin layer. Therefore, the properties of micro and macro-chamber layers were again adjusted at each iteration to optimise their individual fits to the experimental data. A total of 71 iterations were required to reach convergence with $\Delta \text{RMS error} < 0.5\%$ of maximum force and $\Delta \text{maximum strain} < 0.02\%$ of maximum strain for determination of hyperelastic material properties.

After determination of the hyperelastic material properties, the viscoelastic parameters of the heel pad sub-layers were estimated. In total, 6 viscoelastic parameters had to be estimated, $G_1$ and $\beta_1$ for each of the three heel pad sub-layers. The model was simplified as suggested by Hajjarian & Nakarni, by adopting identical decay constants for the three heel pad sub-layers [28]. A similar procedure to that used to obtain the hyperelastic material properties was followed (Figure 4) by fitting the FE predicted results to the corresponding experimental force-strain data but now using the data from the rapid compression tests (225mm/s). Two RMS force errors (one during loading and another during unloading) were used to assess the quality of the model fit. This process was repeated until the errors
did not change significantly with further adjustment (ΔRMS force errors <0.7% of maximum force).

Table 2 shows the result of optimisation at each stage for the heel pad sub-layers.

**Table 2**

Optimisation stages for hyperelastic and viscoelastic models of heel pad sub-layers.

<table>
<thead>
<tr>
<th>Hyperelastic model</th>
<th>First Iteration</th>
<th>Final Iteration</th>
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</thead>
<tbody>
<tr>
<td></td>
<td>µ (kPa)</td>
<td>α (-)</td>
</tr>
<tr>
<td>One layer model</td>
<td>16.45</td>
<td>6.8</td>
</tr>
<tr>
<td>Two layer model</td>
<td>41</td>
<td>4.2</td>
</tr>
<tr>
<td>Micro-chamber</td>
<td>41</td>
<td>4.2</td>
</tr>
<tr>
<td>Macro-chamber</td>
<td>41</td>
<td>4.2</td>
</tr>
<tr>
<td>Skin</td>
<td>104</td>
<td>4.7</td>
</tr>
<tr>
<td>Micro-chamber</td>
<td>104</td>
<td>4.7</td>
</tr>
<tr>
<td>Macro-chamber</td>
<td>35</td>
<td>4.9</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Viscoelastic model</th>
<th>G (MPa)</th>
<th>β ms⁻¹</th>
<th>RMS error loading (%)</th>
<th>RMS error unloading (%)</th>
<th>G (MPa)</th>
<th>β ms⁻¹</th>
<th>RMS error loading (%)</th>
<th>RMS error unloading (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>One layer model</td>
<td>0.39</td>
<td>1</td>
<td>10.1</td>
<td>5.4</td>
<td>0.11</td>
<td>0.08</td>
<td>8.4</td>
<td>5.1</td>
</tr>
<tr>
<td>Two layer model</td>
<td>0.11</td>
<td>0.08</td>
<td>13.4</td>
<td>6.7</td>
<td>0.46</td>
<td>0.06</td>
<td>10.1</td>
<td>4.3</td>
</tr>
<tr>
<td>Micro-chamber</td>
<td>0.11</td>
<td>0.08</td>
<td>11.9</td>
<td>3.5</td>
<td>0.14</td>
<td>0.06</td>
<td>8.6</td>
<td>2.0</td>
</tr>
<tr>
<td>Macro-chamber</td>
<td>0.46</td>
<td>0.06</td>
<td>17.8</td>
<td>7.7</td>
<td>0.42</td>
<td>0.12</td>
<td>17.1</td>
<td>1.8</td>
</tr>
<tr>
<td>Three layer model</td>
<td>0.46</td>
<td>0.06</td>
<td>13.4</td>
<td>12.7</td>
<td>0.30</td>
<td>0.12</td>
<td>14.4</td>
<td>6.4</td>
</tr>
<tr>
<td>Skin</td>
<td>0.14</td>
<td>0.06</td>
<td>13.8</td>
<td>4.1</td>
<td>0.24</td>
<td>0.12</td>
<td>14.5</td>
<td>3.1</td>
</tr>
<tr>
<td>Micro-chamber</td>
<td>0.14</td>
<td>0.06</td>
<td>13.8</td>
<td>4.1</td>
<td>0.24</td>
<td>0.12</td>
<td>14.5</td>
<td>3.1</td>
</tr>
</tbody>
</table>

2.4 Validation

The loaded thickness of the heel pad measured from MRI and the peak plantar pressure under the heel were used to validate the hyperelastic FE model. The loaded MRI was taken from the right foot of the subject whose unloaded MRI data was used previously to build the heel pad model. A device was developed to load and vertically compress the heel pad during MRI scanning. The load and compression mimicked the loading in the STRIDE and the FE model. The device comprised of a
wooden foot support under the heel (rotated by ~17° into dorsiflexion) attached to a harness worn by the subject during scanning. Elastic straps attaching the harness to the footplate were adjusted to create tension and thus compress the heel (Figure 5).

Figure 5. The heel pad loading device. \( L = \) force applied to plantar aspect of heel.

The applied load and pressure were measured using a Pedar pressure measurement insole system with a resolution of 2.5-5 kPa (Novel.de, Munich, Germany) before entering the MRI scanner. Some pilot measurements were performed before and after MRI scanning to ensure that using the heel pad loading device provides consistent data out of and during MRI scanning. The force was measured for 17 sensors with total area of 3295mm² under the heel region. Larger insole than the foot size was selected to ensure that not any load or pressure data of the heel is missed. The T1 MRI scans were taken with 160×160 pixels and spacing of 5.5mm from the heel area in the coronal view. During the MRI scanning, the subject was lying in the supine position. The loaded thickness was measured at the image slice 29mm from the back of the heel, which was closest to the calcaneus tuberosity, and 34mm from the lateral side. To predict the loaded thickness of the heel pad and plantar pressure in the FE model, the indenter and load cell were replaced by a rectangular flat rigid plate.
To demonstrate that the viscoelastic FEA model could extrapolate from the results used to find the material properties, different experimental results were used for validation, including results for rapid compression tests at 141mm/s and sinusoidal loading. RMS errors between force-strain responses of the heel pad during loading and unloading periods were used to evaluate the quality of the viscoelastic FE model in reproducing the behaviour of the heel pad at rapid compression tests.

3. Results

Using inverse FEA, hyperelastic and viscoelastic material properties were obtained for the macro-chamber, micro-chamber and skin layers (Figure 6 and Tables 3, 4). In Figure 6 visual inspection of the graphs confirms that the heel pad and its sublayers show nonlinear behaviour under loading. For a 36.5% strain of the heel pad under slow compression, macro-chamber and micro-chamber strains were 41.8 and 25.3% respectively. These values were 41.7 and 26.3% for macro-chamber and micro-chamber respectively, under rapid compression. During the hold period while the displacement was kept constant, the load decreased illustrating the stress-relaxation characteristics of the tissue layers. During unloading the heel pad and indenter lost contact around 20% strain. This can be explained by the fact that the heel pad returned to its original shape at a slower rate than the indenter velocity. In viscoelastic modelling, the maximum error was obtained at the middle portion of the loading period where the Ogden material model was not able to simulate the nonlinear behavior of the tissue accurately.
Figure 6. Macro-chamber, micro-chamber and heel pad behaviour under slow and rapid compression (data used for material properties estimation).

Table 3
Final hyperelastic material properties of the heel pad sub-layers.

<table>
<thead>
<tr>
<th></th>
<th>µ (MPa)</th>
<th>α (-)</th>
<th>RMS force error (N)</th>
<th>Difference between max strains</th>
</tr>
</thead>
<tbody>
<tr>
<td>Skin</td>
<td>0.452</td>
<td>5.6</td>
<td>1.98 (2.7%)</td>
<td>-</td>
</tr>
<tr>
<td>Micro-chamber</td>
<td>0.095</td>
<td>4.9</td>
<td>3.73 (5.0%)</td>
<td>0.3%</td>
</tr>
<tr>
<td>Macro-chamber</td>
<td>0.036</td>
<td>4.5</td>
<td>1.92 (2.6%)</td>
<td>0.4%</td>
</tr>
</tbody>
</table>

Table 4
Final viscoelastic material properties of the heel pad sub-layers.

<table>
<thead>
<tr>
<th></th>
<th>G₁ (MPa)</th>
<th>β₁ (milli-seconds)¹</th>
<th>RMS force error Loading (N)</th>
<th>RMS force error Unloading (N)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Skin</td>
<td>0.42</td>
<td>0.12</td>
<td>19.88 (17.1%)</td>
<td>2.15 (1.8%)</td>
</tr>
<tr>
<td>Micro-chamber</td>
<td>0.30</td>
<td>0.12</td>
<td>16.74 (14.4%)</td>
<td>7.44 (6.4%)</td>
</tr>
<tr>
<td>Macro-chamber</td>
<td>0.24</td>
<td>0.12</td>
<td>16.91 (14.5%)</td>
<td>3.59 (3.1%)</td>
</tr>
</tbody>
</table>
The hyperelastic model predicted the loaded (~315N) heel pad thickness within 6.4% of the thickness measured via MRI. The hyperelastic model showed similar peak plantar pressure compared to the experimental data from Pedar system. Figure 7 compares the numerical and experimental results of the plantar pressure under 315N. As shown by the contour plot of the numerical result, the peak pressure appeared in the central region of the heel with the value 215kPa (averaged over the area of 10x19mm² that is close to the one sensor size in the Pedar insole). This is comparable with the results of Pedar system measurements with the value of 225kPa at the very similar location with error of 4.4% maximum peak plantar pressure.
Figure 7. Validation of hyperelastic model under compressive load of 315N: (A) Loaded MR image of the heel pad; (B) FE model of the loaded heel pad; (C) Pedar pressure insole measurement; (D) FE model pressure prediction

The viscoelastic model could simulate the heel pad behaviour with RMS force errors of 13.8-14.7% and 1.6-5.2% of maximum force for loading and unloading periods respectively for rapid loading (141mm/s). The viscoelastic model simulated the heel pad behavior under sinusoidal loading with RMS force errors of 4.5-8.9% and 2.6-5.8% of maximum force for loading and unloading periods respectively.

4. Discussion

The initial elastic modulus of the macro-chamber, micro-chamber and skin layers was 0.243, 0.698 and 3.797MPa, respectively. Direct comparisons to prior literature are difficult because there are no previous reports of the three separate layers. Like our study, Hsu et al. used in-vivo data and their elastic modulus of 0.181MPa for the macro-chamber layer concurs quite well with the value reported here (0.243MPa) [29]. Their micro-chamber layer modulus was 1.140MPa, almost twice the stiffness reported here (0.698MPa). This is probably due to Hsu et al. combining the micro and much stiffer skin layers together resulting in an apparent elevation in micro-chamber layer stiffness. Erdemir et al. reported a much lower elastic modulus of 0.050MPa (SD 0.025) for a homogenous heel pad (i.e. all three layers combined) using inverse FEA and in-vivo experimental data for 20 subjects [7]. The values obtained here are outside their range and this is perhaps due to their use of an axisymmetric rectangular heel pad model and compression system (a 25.4mm diameter indenter). Using an inverse FEA method, a value of 0.300MPa was reported from impact testing of isolated heel tissue [11], which is comparable with the value for the macro-chamber layer reported here (0.243MPa). In another case, elastic moduli of 0.003 and 6.528MPa were derived for the heel fat pad and skin respectively, based on in-vitro and in-vivo experimental data [8]. The value for the skin layer of 6.528MPa is much higher than the value reported here (3.797MPa), but clearly represents a layer far stiffer than macro and micro layers. The difference is likely due to the small value of 0.003MPa they found for the fat pad based on experimental data from unconfined testing of isolated fat samples.
Clearly, different initial elastic moduli have been reported for the heel pad and its sub-layers and data are sensitive to the choice of experimental methods (*in-vivo* or *in-vitro*), age and health of subjects/samples, number of subjects/samples and the degree of simplification of the model used for inverse FEA.

Because of variations in material model definitions of $\alpha$ (deviatoric exponent), the values reported here should only be compared to studies which used the same model. In this study $\alpha$ was 4.5, 4.9 and 5.6 for the macro-chamber, micro-chamber and skin layers respectively. Erdemir et al. reported a value of 6.82 (SD 1.57) for the entire heel pad [7]. Values of 8.8 and 6.8 have been reported for the fat pad and skin, respectively [8]. It is difficult to judge the appropriateness of direct comparisons since so few participants are used in these experiments and models.

A time constant of ~8ms (reciprocal of the decay constant $\beta$) was found for all heel pad sub-layers. Values of 1 and 2ms were reported from inverse FEA using compression data of cadaveric intact heel pads [12, 30]. However, they used experimental data collected from a different foot than that used to build the model geometry. In another case, the time constant was 500ms for the heel pad (from experiments on dissected fat pad samples) [13]: a result that may have been affected by dehydration of the sample.

The relaxation modulus is represented differently in different FEA software making comparisons difficult. While Ls-Dyna uses shear relaxation modulus ($G_1$), ABAQUS uses relaxation coefficient ($g$) which is equal to $G_1/G\infty+G_1$. $G\infty$ is the long-term shear modulus and it is $\geq \frac{1}{3}$ of the initial elastic modulus. Based on the above relations, $0 \leq g \leq 1$ and when $g \to 1$ the material shows characteristics that are more viscoelastic and when $g \to 0$ the material shows characteristics that are more elastic. Having the initial elastic moduli of the heel pad sub-layers ($3.797$, $0.698$ and $0.243$MPa), $g$ was estimated as

$$ g = \begin{cases} 0.42 \frac{0.42}{G_{\infty} \geq 1.26} + 0.42 & \leq 0.25, \\ 0.30 \frac{0.30}{G_{\infty} \geq 0.23} + 0.30 & \leq 0.57, \\ 0.24 \frac{0.24}{G_{\infty} \geq 0.08} + 0.24 & \leq 0.75 \end{cases} $$

for the skin, micro-chamber and macro-chamber respectively. In this study, $g$ complies with the general rule (i.e. is between 0 and 1) and from skin to macro-chamber, the viscoelastic behaviour of the materials increases. Previously $g$ was reported as 0.99 using inverse FEA [12, 30], representing a highly viscous heel pad.
The 3-layer heel pad model reported here predicted static heel pad thickness under load with an error of 6.4%, which is towards the lower end of the range (5-15%) reported previously for 1-layer and 2-layer models [7, 8, 9, 10]. Spears et al. showed that while a 1-layer model overestimated the plantar heel pressure at the centre of the model (>60% error) and underestimated it at medial and lateral regions (>100% error), a 2-layer model predicted pressures far closer to experimental data (within 10%) [8]. Similarly the 3-layer model reported here produced even small error (<4.4% maximum peak plantar pressure); suggesting that the additional increase in model complexity was justified.

Unique to this study, the unloading behaviour of the model was evaluated in addition to the behaviour during loading. The errors propagated during the estimation of material properties of the three heel pad sub-layers might be the source of this error. The measurement of the applied load under the heel using Pedar system might be another source of these errors.

Since the study included one participant, all findings are unique to the properties of the particular heel studied. It is acknowledged that the inverse FEA process used to determine the material properties, which is based on a manual search procedure, might find different local minima when different initial values for the material properties are used. To simplify the model, identical time constants were used for all three layers. However, given that other properties differ significantly between the layers, this simplification may not be appropriate. All results were obtained for compression loading at a single angle of rotation of the heel and in the absence of shear loading.

To our knowledge, this is the first study to estimate the hyperelastic and viscoelastic material properties of the heel pad sub-layers using in-vivo data and loading conditions similar to those experienced during gait and standing. Like other FE models, not only can this model predict pressures and shear stresses at the plantar surface but it can also be used to predict internal tissue mechanics.

Future work using the model could include studies of the effects of footwear materials on internal stresses and the effects of experimental conditions on heel pad behaviour (such as indenter size and shape).

Conflict of Interest
The authors have no financial or personal relationships with other people or organisations that could inappropriately influence this work.

References


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