



University of
Salford
MANCHESTER

Deformed wing virus is a recent global epidemic in honeybees driven by Varroa mites

Wilfert, L, Long, G, Leggett, H, Schmid-hemple, P, Butlin, R, Martin, SJ and Boots, M

<http://dx.doi.org/10.1126/science.aac9976>

Title	Deformed wing virus is a recent global epidemic in honeybees driven by Varroa mites
Authors	Wilfert, L, Long, G, Leggett, H, Schmid-hemple, P, Butlin, R, Martin, SJ and Boots, M
Type	Article
URL	This version is available at: http://usir.salford.ac.uk/id/eprint/38152/
Published Date	2016

USIR is a digital collection of the research output of the University of Salford. Where copyright permits, full text material held in the repository is made freely available online and can be read, downloaded and copied for non-commercial private study or research purposes. Please check the manuscript for any further copyright restrictions.

For more information, including our policy and submission procedure, please contact the Repository Team at: usir@salford.ac.uk.

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31

Title Deformed Wing Virus is a Recent Global Epidemic in Honeybees driven by Varroa Mites

Authors: L. Wilfert^{1*}, G. Long², H. C. Leggett^{2†}, P. Schmid-Hempel³, R. Butlin², S.J.M. Martin^{2‡}, M. Boots^{1§}.

Affiliations:

¹Centre for Ecology and Conservation, University of Exeter, Penryn Campus, Penryn TR11 9FE, UK.

²Department of Animal and Plant Sciences, University of Sheffield, Sheffield, S10 2TN, UK.

³Institute of Integrative Biology, ETH Zürich, 8092 Zürich, Switzerland.

*Correspondence to: lena.wilfert@ex.ac.uk

†Department of Genetics, University of Cambridge, Cambridge, CB2 3EH, UK.

‡School of Environment and Life Sciences, University of Salford, Manchester, M5 4WT, UK.

§Department of Integrative Biology, University of California, Berkeley, CA, 94720, USA.

Abstract: Deformed Wing Virus (DWV) and its vector *Varroa destructor*, which emerged last century, are a major threat to the world's honeybees. While *Varroa*'s dramatic impacts on colony-level DWV epidemiology is evident, we have little understanding of wider DWV epidemiology and the role that *Varroa* has played in its global spread. A phylogeographic analysis shows that DWV is globally distributed in honeybees, having recently spread from a common source, the European honeybee *Apis mellifera*. DWV shows epidemic growth and transmission that is predominantly mediated by European and North American honeybee populations and driven by trade and movement of honeybee colonies. DWV is now an important re-emerging pathogen of honeybees undergoing a worldwide man-made epidemic, fuelled by the novel direct transmission route provided by the *Varroa* mite.

One Sentence Summary: Honeybees are undergoing a DWV pandemic, coinciding with the emergence of the *Varroa* mite, with the global spread driven by Western bee populations.

32 **Main Text:** The European honeybee *Apis mellifera* can be argued to be one of the most
33 important domesticated animals, heavily used for commercial pollination of intensive and high-
34 value crops such as the California almond, macadamia, cherries or blueberries as well as honey
35 production. *A. mellifera*, originally from East Asia (1), has been intensively managed by
36 beekeepers and exported from its native population in Europe and Africa to the New World and
37 Oceania by European settlers, where beekeeping has become widespread in the last century in
38 line with agricultural intensification. Although wild pollinators play an important role not only
39 for wild flowering plants but also for crop pollination (2), our current horticultural systems now
40 heavily rely on managed honeybees, and the global stock of domesticated honeybees is growing
41 more slowly than agricultural demand for pollination (3). Understanding the key threats to *A.*
42 *mellifera* is, as a consequence, clearly important if we are to maintain large populations of bees
43 for both honey production and pollination services. While the number of honeybee hives has
44 increased by 45% on a global scale, there have been dramatic regional declines (e.g. a reduction
45 of 59% in the USA from 1947 to 2005) and beekeepers now globally report high over-wintering
46 colony mortalities, which threaten their sustainability (4). While many factors ranging from
47 agricultural intensification to the use of pesticides have been implicated in pollinator declines
48 (5), RNA viral infections vectored by the ectoparasitic mite *Varroa destructor* have the potential
49 to be major contributors to global honeybee colony mortalities (6). In particular, Deformed Wing
50 Virus (DWV) is the key pathogen associated with over-winter mortality of *Varroa*-infested
51 colonies (7-10). The *Varroa* mite jumped from its native host, the Asian honeybee *A. cerana*, to
52 the European honeybee, *A. mellifera*, in the middle of the last century and now has a global
53 distribution (11). While DWV occurs in *Varroa*-free natural populations (12-14), DWV
54 replicates in the mite (15, 16) or potentially accumulates in its gut ((17), but see (18)). *Varroa*
55 can inject the virus directly into the bee's hemolymph (15, 19), thus circumventing some of the
56 natural infection barriers to vertical or horizontal transmission between bees, such as the
57 exoskeleton and the peritrophic membranes lining the digestive tract (20). Indeed, the recent
58 *Varroa* invasions in Hawaii (12) and New Zealand (13) led to an increase in DWV prevalence
59 both across colonies and in the viral load in infected individuals, coinciding with a loss in viral
60 diversity. These natural experiments (12, 13) have demonstrated that *Varroa* increases the spread
61 of DWV in honeybee populations. There is also evidence that *Varroa* not only acts as a vector
62 but also increases the virulence of DWV infections, turning relatively asymptomatic infections

63 into ‘overt’ infections associated with clinical disease symptoms (15, 21-23) and increasing
64 winter colony mortalities (7-10). There is therefore good evidence that *Varroa* impacts
65 individual and colony-level DWV epidemiology in honeybees, but its importance to the global
66 spread and ongoing worldwide transmission of DWV is unknown. This is an important problem
67 because honeybees today have both a global distribution and a global market. Therefore, we need
68 to understand the factors that drive disease transmission on a global scale in order to be able to
69 limit the spread of the pathogen and mitigate negative effects on beekeeping and the ecosystem
70 services provided by bees (4). Furthermore, honeybee diseases also impact the wider pollinator
71 community (24, 25) and we need to understand the global drivers of disease spread to manage
72 disease transfer to novel hosts.

73
74 Here, we use a phylogeographic approach to test whether *Varroa*-vectored DWV is a globally
75 emerging honeybee pathogen and to determine the dominant routes of DWV spread. There are
76 two main scenarios for DWV’s origin that can be distinguished based on its phylogeography.
77 The first scenario is that *Varroa* introduced DWV to the European honeybee *A. mellifera* and
78 caused a global epidemic. Under this scenario, we would expect East-Asian *Varroa* populations
79 to be the ancestral host of DWV. The second scenario is that DWV is a re-emerging disease
80 whose current pandemic is promoted by *Varroa*, in which case we would expect *A. mellifera* as
81 the ancestral host. We estimate the major routes of global transmission by comparing geographic
82 and host-specific patterns dated via the viral evolutionary rate, which we have derived for three
83 genomic fragments. A total of 246 DWV sequences were collected from honeybees and *Varroa*
84 mites in thirty-two geographic locations in seventeen countries world-wide, supplemented by all
85 publicly available DWV sequence data, and used to infer the epidemic and migration history
86 driving present-day global DWV dynamics.

87
88 From our analysis, DWV shows a recent global radiation and pandemic, with the most recent
89 common ancestor coinciding in time with the global emergence of the *Varroa* mite as a
90 honeybee ectoparasite in the middle of the last century (11). The most recent common ancestor
91 for each fragment dates back to the middle of the last century with mean root heights of 44 years
92 (*rdrp*-fragment, 95% Highest Posterior Density (HPD) 27 - 63 years), 47 years (*vp3*-fragment, 95
93 % HPD 28 – 74 years) and 78 years (*lp*-fragment, 95 % HPD 45 -118 years). All fragments show

94 significant exponential growth over the last decades, with doubling rates around 13 years [*lp*-
95 fragment 16.4 years (95% HPD 9.9 – 46.8 years), *rdrp*-fragment 11.6 years (95% HPD 6 – 96.6
96 years), *vp3*-fragment 12.4 years (95 % HPD 6.1 – 262.8 years)], which is supported by GMRF
97 skyride analysis (supplementary Fig. S4). Since population structure tends to produce a spurious
98 signature of declining effective population sizes (26), we excluded the small number of
99 geographically disparate samples available from Genbank from 2010 for demographic analyses
100 (see Database S1). With the exception of the *rdrp*-fragment, exponential growth is also
101 significant when including samples from 2010-2013. In combination, these results lend support
102 to the hypothesis that DWV has recently radiated from a common source and exponentially
103 spread across the globe (27).

104
105 While this demographic pattern is consistent with an important temporal role for *Varroa* in the
106 recent expansion of DWV, the global distribution and the ancestral host state of this virus is
107 consistent with DWV being a re-emerging honeybee virus. DWV has been isolated from
108 honeybee populations that had not been exposed to *Varroa* (Australia ((28) (HQ655496-
109 HQ655501) and present study, see also Fig. S5), Colonsay Island (Scotland) (14), Hawaii (12),
110 Ile d'Ouessant (France) (14), Isle of Man (present study), Newfoundland (29) and New Zealand
111 (13)). This alone would not preclude *Varroa* as the initial source for DWV in *A. mellifera*, as
112 novel emerging pathogens can spread ahead or independently of the initial host if they can
113 replicate in their novel host, as is the case not only in many human zoonoses, such as SARS, but
114 also in wildlife diseases, such as squirrel pox (30, 31). Here, *Varroa*, as an active vector that
115 increases DWV prevalence and titer in honeybees (12, 13), may increase human-mediated viral
116 spread by increasing the number of infected bees and their transmission potential even without
117 the mite being spread itself. In addition to DWV-presence in *Varroa*-free populations, the
118 phylogenetic reconstruction also contradicts *Varroa* as the ancestral host of the virus. The
119 ancestral host is unanimously identified as *A. mellifera* (state probability $P_{lp} = 99.43\%$, $P_{vp3} =$
120 97.18% , $P_{rdrp} = 92.7\%$) – not *V. destructor* (Fig. 1) nor *A. cerana* (Fig. S6 and S7). The
121 geographic origin is less certain with ancestral states being reconstructed with low probabilities,
122 (*lp*-fragment: East Asia, $P_{lp} = 69.77\%$, *vp3*- and *rdrp*-fragments: Pakistan, $P_{vp3} = 77.25\%$, $P_{rdrp} =$
123 54.84%). While we cannot categorically rule out that DWV was introduced to honeybees from
124 an entirely unknown host, this pattern rules out *Varroa* as well as *A. cerana* as the ancestral

125 DWV-host. The most parsimonious explanation for this pattern is our second scenario: DWV is
126 an endemic honeybee pathogen that has recently re-emerged through ecological change, the
127 spread of *Varroa* as a vector, alongside increased global movement of infected bees or other
128 material such as pollen. This supports previous work postulating that the ancestral form of DWV
129 may have been associated with *A. mellifera* (32) and that similarities between DWV lineages
130 may represent a recent introduction from *A. mellifera* into other *Apis* species (33).

131
132 Our data show that the recent spread of DWV is driven by European *A. mellifera* populations
133 (Fig. 1 & 2a) and shows a similar pattern to the spread of *Varroa* (Fig. 2b), despite increased
134 regulation and control of the global trade in honeybees (11). Combining results from the three
135 fragment subsamples for the DWV subtype, Europe, followed by North America, emerge as the
136 main hubs of transmission for DWV to the New World (North and South America and Hawaii)
137 and Oceania (Australia and New Zealand) (Fig. 2 and supplementary table S5). Additionally,
138 there is strong support for migration between East Asia and Europe, with migration being
139 supported in both directions, as well as from Pakistan to Europe in the case of the *vp3*- and *rdrp*-
140 fragments. This pattern overall reflects the invasion pattern of the *Varroa* mite (Fig. 2). Small
141 differences in migration patterns between the fragments may be caused by real biological
142 differences: DWV shows evidence of frequent recombination (15) and thus genes may differ in
143 their evolutionary history as well as in their evolutionary rate. However, these differences can
144 also potentially be explained by the different subsets of samples available across fragments
145 (Table S4). Additional analyses to address unequal sample distribution and a sampling bias
146 towards European populations confirmed the predominant pattern of European and North
147 American populations as the main transmission hubs, with some evidence for transmission from
148 Asia to these hubs (Table S6). This analysis also shows strong support for transmission from *A.*
149 *mellifera* to *V. destructor* for all fragments (Bayes Factor $BF_{lp}=12281.21$, $BF_{vp3}=1813.53$,
150 $BF_{rdrp}=12281.21$) as well as to other hosts (the common Asian honeybee ectoparasite
151 *Tropilaelaps calreae*, *lp*-fragment $BF = 11051.99$, and the bumblebee *Bombus lapidarius*, *rdrp*-
152 fragment $BF = 4.62$) as shown in Fig. 3. These are not dead-end hosts, with limited evidence for
153 transmission to *A. mellifera* (*V. destructor* to *A. mellifera*: $BF_{lp}=3.97$, $BF_{vp3}=1813.53$,
154 $BF_{rdrp}=3.09$; *rdrp*-fragment: *B. lapidarius* to *A. mellifera* $BF=3.74$, *lp*-fragment: *T. clareae* to *A.*
155 *mellifera* $BF=3.93$). DWV shows very little host specificity, as the viral population is not

156 structured by host species: K_{ST} , which measures the proportion of genetic variation among
157 populations, is non-significant or close to zero ($K_{ST_{lp}} = 0.023$, $K_{ST_{rdrp}} = 0.02$, both $p < 0.05$,
158 $K_{ST_{vp3}}$ n.s.). In contrast, there is significant but overall moderate geographic population
159 differentiation for all fragments ($K_{ST_{lp}} = 0.305$, $K_{ST_{vp3}} = 0.703$, $K_{ST_{rdrp}} = 0.422$, all $p < 0.001$).
160 Population differentiation is significant, but less pronounced within Europe ($K_{ST_{lp}} = 0.319$,
161 $K_{ST_{vp3}} = 0.135$, $K_{ST_{rdrp}} = 0.181$, all $p < 0.001$) and East Asia ($K_{ST_{lp}} = 0.301$, $p < 0.001$; other
162 areas/fragments provided too few samples to be informative). Samples that are genetic nearest
163 neighbors largely come from the same population (Hudson's nearest neighbor statistic at
164 continent level: $S_{nn_{lp}} = 0.831$, $S_{nn_{vp3}} = 0.679$, $S_{nn_{rdrp}} = 0.65$, all $p < 0.001$; within Europe: $S_{nn_{lp}}$
165 $= 0.772$, $S_{nn_{vp3}} = 0.771$, $S_{nn_{rdrp}} = 0.628$, both $p < 0.001$; within East Asia: $S_{nn_{lp}} = 0.923$, $p <$
166 0.001). This indicates that DWV has accrued geographic variation since the origin of the
167 epidemic ~80 years ago, but highlights that high rates of human-mediated migration within
168 Europe and East Asia may obscure population differentiation. It is also evident from the
169 phylogenetic trees (Fig. 1) that *A. mellifera* is the reservoir host for DWV, with other host
170 species clustered at the terminal nodes. Thus DWV apparently has little host specificity, being
171 readily transmitted between different host species, but its primary host is *A. mellifera*, with
172 global transmission having largely been driven by European populations (Fig. 2).

173
174 DWV not only causes colony mortality in managed *A. mellifera* populations but also impacts
175 feral populations (34) and has been identified as an emerging disease in wild pollinators (24, 25,
176 35), with dramatic impacts on survival in bumblebees (24). As such DWV may pose a threat not
177 only to managed honeybees but also to pollinators more generally. Wild pollinators such as
178 bumblebees and solitary bees have experienced a loss of species richness and diversity over the
179 last decades, which can partly be attributed to infectious diseases (4, 36-39). Our results show
180 that there is a global pandemic of DWV with transmission mediated by European populations of
181 *A. mellifera*. This is an anthropogenic transmission, spread by the global movement of honeybees
182 or other infected material, likely fueled by the concurrent emergence of *V. destructor* mites. This
183 highlights how pollinator populations are globally inter-connected via the trade and movement of
184 managed pollinators, leading to the rapid potential spread of pathogens and parasites across the
185 globe and between species. To control DWV and to reduce the negative effects of DWV on
186 beekeeping and wild pollinators, tighter controls such as mandatory health screening and

187 restricted movement of honeybees across borders should be imposed, with every effort made to
188 maintain the current *Varroa*-free refugia for the conservation of wild and managed pollinators in
189 the absence of this vector.

190 **References and Notes:**

- 191 1. A. Wallberg *et al.*, *Nature Genetics* **46**, 1081-1088 (2014).
192 2. L. A. Garibaldi *et al.*, *Science* **339**, 1608-1611 (2013).
193 3. M. A. Aizen *et al.*, *Curr Biol* **19**, 915-918 (2009).
194 4. S. G. Potts *et al.*, *Trend Ecol Evol* **25**, 345-353 (2010).
195 5. A. J. Vanbergen *et al.*, *Frontiers Ecol Envir* **11**, 251-259 (2013).
196 6. E. Genersch *et al.*, *Vet Res* **41**, (2010).
197 7. A. C. Highfield *et al.*, *Appl Environ Microbiol* **75**, 7212-7220 (2009).
198 8. H. Berthoud *et al.*, *J Api Res* **49**, 60-65 (2010).
199 9. B. Dainat *et al.*, *Appl Env Microbiol* **78**, 981-987 (2012).
200 10. E. Genersch *et al.*, *Apidologie* **41**, 332-352 (2010).
201 11. B. P. Oldroyd, *Trend Ecol Evol* **14**, 312-315 (1999).
202 12. S. J. Martin *et al.*, *Science* **336**, 1304-1306 (2012).
203 13. F. Mondet *et al.*, *PLoS Pathog* **10**, (2014).
204 14. C. Mouret *et al.*, *Rev Met Vet* **164**, 577-582 (2013).
205 15. E. V. Ryabov *et al.*, *PLoS Path* **10**, (2014).
206 16. S. Gisder *et al.*, *J Gen Virol* **90**, 463-467 (2009).
207 17. T. Erban *et al.*, *Sci Rep* **5**, (2015).
208 18. D. Cardoen *et al.*, *PLoS ONE* **6**, (2011).
209 19. J. R. de Miranda *et al.*, *J Invert Pathol* **103**, S48-S61 (2010).
210 20. J. D. Evans *et al.*, *J Invert Pathol* **103**, S62-S72 (2010).
211 21. P. L. Bowen-Walker *et al.*, *J Invert Pathol* **73**, 101-106 (1999).
212 22. S. J. Martin, *J Appl Ecol* **38**, 1082-1093 (2001).
213 23. C. Yue *et al.*, *J Gen Virol* **86**, 3419-3424 (2005).
214 24. M. A. Fürst *et al.*, *Nature* **506**, 364-366 (2014).
215 25. R. Manley *et al.*, *J Appl Ecol* **52**, 331-340 (2015).
216 26. R. Heller *et al.*, *PLoS ONE* **8**, e62992 (2013).
217 27. O. Berenyi *et al.*, *Appl Env Microbiol* **73**, 3605-3611 (2007).
218 28. R. Singh *et al.*, *PLoS ONE* **5**, (2010).
219 29. D. Shutler *et al.*, *PLoS ONE* **9**, (2014).
220 30. D. M. Tompkins *et al.*, *Proc R Soc B* **269**, 529-533 (2002).
221 31. D. M. Tompkins *et al.*, *Ecol Lett* **6**, 189-196 (2003).
222 32. X. Zhang *et al.*, *J Invert Pathol* **109**, 156-159 (2012).
223 33. J. Li *et al.*, *PLoS ONE* **7**, (2012).
224 34. C. E. Thompson *et al.*, *PLoS ONE* **9**, (2014).
225 35. D. P. McMahon *et al.*, *J Anim Ecol* **84**, 615-624 (2015).
226 36. J. C. Biesmeijer *et al.*, *Science* **313**, 351-354 (2006).
227 37. S. A. Cameron *et al.*, *PNAS* **108**, 662-667 (2011).
228 38. L. G. Carvalheiro *et al.*, *Ecol Lett* **16**, 1416-1417 (2013).
229 39. J. Ollerton *et al.*, *Science* **346**, 1360-1362 (2014).
230 40. E. Forsgren *et al.*, *Exp Appl Ecol* **47**, 87-97 (2009).
231 41. G. Lanzi *et al.*, *J Virol* **80**, 4998-5009 (2006).
232 42. E. Genersch, *Vet J* **169**, 121-123 (2005).
233 43. S. L. K. Pond *et al.*, *Bioinformatics* **21**, 676-679 (2005).
234 44. D. P. Martin *et al.*, *Bioinformatics*, (2010).

235 45. J. Felsenstein. (Distributed by the author. Department of Genome Sciences, University of
236 Washington, Seattle., 2005).

237 46. S. Alizon *et al.*, *Retrovirology* **10**, (2013).

238 47. C. Ramsden *et al.*, *Mol Biol Evol* **26**, 143-153 (2009).

239 48. G. Baele *et al.*, *Mol Biol Evol* **29**, 2157-2167 (2012).

240 49. G. Baele *et al.*, *Mol Biol Evol* **30**, 239-243 (2013).

241 50. A. J. Drummond *et al.*, *Bayesian evolutionary analysis with BEAST 2*. (Cambridge
242 University Press, Cambridge, 2014).

243 51. D. Posada, *Mol Biol Evol* **25**, 1253-1256 (2008).

244 52. R. R. Gray *et al.*, *BMC Evol Biol* **11**, (2011).

245 53. A. J. Drummond *et al.*, *Genetics* **161**, 1307-1320 (2002).

246 54. V. N. Minin *et al.*, *Mol. Biol. Evol.* **25**, 1459-1471 (2008).

247 55. A. L. Hicks *et al.*, *J Virol* **85**, 7942-7947 (2011).

248 56. P. Lemey *et al.*, *PLoS Comput Biol* **5**, e1000520 (2009).

249 57. M. A. R. Ferreira *et al.*, *Canad. J. Stat.* **36**, 355-368 (2008).

250 58. R. Kajobe *et al.*, *J Invert Pathol* **104**, 153-156 (2010).

251 59. E. Muli *et al.*, *PLoS ONE* **9**, (2014).

252 60. N. J. Haddad *et al.*, *Insect science*, (2015).

253 61. N. De Maio *et al.*, *PLoS Genetics* **11**, (2015).

254 62. P. Librado *et al.*, *Bioinformatics* **25**, 1451-1452 (2009).

255 63. R. R. Hudson *et al.*, *Mol. Biol. Evol.* **9**, 138-151 (1992).

256 64. R. R. Hudson, *Genetics* **155**, 2011-2014 (2000).

257 65. S. L. Arlinghaus *et al.*, *Solstice* **9**, (1998).

258 66. V. Dietemann *et al.*, *Apidologie* **40**, 285-295 (2009).

259 67. D. Begna, *J Fish Livestock Prod* **3**, (2014).

260 68. M. H. Allsopp, University of Pretoria, (2006).

261 69. E. Crane, *The world history of beekeeping and honey hunting*. (Routledge, New York,
262 1999).

263 70. J. Lindsay, in *Proceedings of the Third Caribbean Beekeeping Congress*, L. E. McLaren,
264 Ed. (2002).

265 71. M. H. Allsopp, in *Sub-regional training seminar on diseases of honey bees for OIE*
266 *national focal points for animal disease notification to the OIE* Y. Samake, Ed. (Oie,
267 Swaziland, 2011).

268 72. C. Davey, *Beekeepers Quart* **100**, 84 (2010).

269 73. L. I. de Guzman *et al.*, *Biochem Genet* **35**, 327-335 (1997).

270 74. S. Gladstone, in *Bees for Development*. (beesfordevelopment.org, 2011), pp. 103-110.

271 75. G. Salamanca *et al.*, *Zootec Trop* **30**, 183-195 (2012).

272 76. K. Akinwande *et al.*, *Afric J Food Agr Nut Dev* **13**, (2013).

273 77. H. Denmark *et al.*, *Ent Circ* **347**, (1991).

274 78. M. El-Niweiri *et al.*, *AlBuhuth* **10**, 61-76 (2006).

275 79. H. Rasolofoaivao *et al.*, *Exp Appl Acar* **60**, 521-530 (2013).

276 80. A. Paraiso *et al.*, *J Api Res* **50**, 321-322 (2011).

277 81. M. Frazier *et al.*, *Apidologie* **41**, 463-465 (2010).

278 82. D. L. Anderson *et al.*, *Exp Appl Acarol* **24**, 165-189 (2000).

279 **Acknowledgments:**

280 Acknowledgements: The data reported in this paper are available on Genbank (Accession
281 numbers KP734326-734846; KP765048-765235), in the Supplementary Material and in
282 Supplementary Database S1. We would like to thank numerous beekeepers for samples (see
283 supplementary material). We are grateful to Andrew Cowley and MD Sharma for informatics
284 support and to Floh Bayer for help in the lab. This work was funded by a NERC grant to MB,
285 SM and RB (NE/F019610/1); LW was funded by a Royal Society Dorothy Hodgkin Fellowship.

286

287 **Fig. 1:** Phylogenetic reconstruction of three fragments of DWV showing host and geographic
288 structure. The figure shows Maximum clade credibility (MCC) trees for the *lp*-fragment (A),
289 *vp3*-fragment (B) and the *rdrp*-fragment (C) of DWV. The branches are colored according to the
290 lineages' inferred geographic origin and the nodes are colored according to the inferred host
291 species. Posterior support >0.5 is indicated for nodes up to the 4th order; horizontal bars indicate
292 the time scale in years. The x-Axis shows time in years. The pie charts show the inferred
293 posterior distribution of the root's geographic location state. See Fig. S3 for an alternative
294 visualization of this graph.

295

296 **Fig. 2:** Global migration patterns of DWV and *V. destructor*. a) Phylogenetically inferred major
297 migration patterns of DWV. The weight of the line indicates the Bayes Factor support for non-
298 zero transition rates (from thin to thick arrows: BF = 3 – 10, 10 – 100, >100) and the color
299 indicates the fragments for which these routes were supported (note that the Thai population was
300 only available for the *lp*-fragment; see Table S5 for detailed results). b) Temporal spread of *V.*
301 *destructor* in *A. mellifera* based on first records per country (see Materials and Methods); to
302 reflect the coarseness in the data, the temporal spread is indicated by decade. Currently, the only
303 remaining *Varroa*-free large land-masses with a significant honey bee population are Australia
304 and Newfoundland, with mounting evidence that sub-Saharan Africa has been invaded since the
305 turn of the century.

306 **Fig. 3** Phylogenetically inferred DWV-host switching patterns. The weight of the line indicates
307 the Bayes Factor support for non-zero transition rates (from thin to thick arrows: BF = 3 – 10, 10
308 – 100, >100) and the color indicates the fragments for which these routes were supported

309

310 **Supplementary Materials:**

311 Materials and Methods

312 Figures S1-S7

313 Tables S1-S6

314 References (40-82)